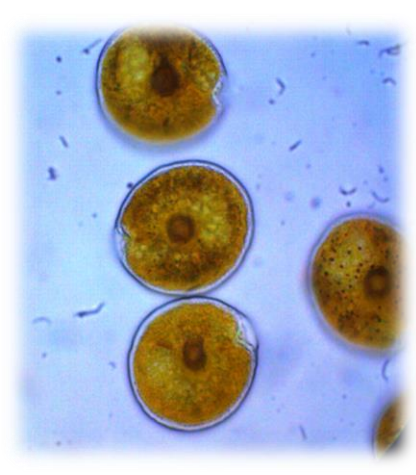
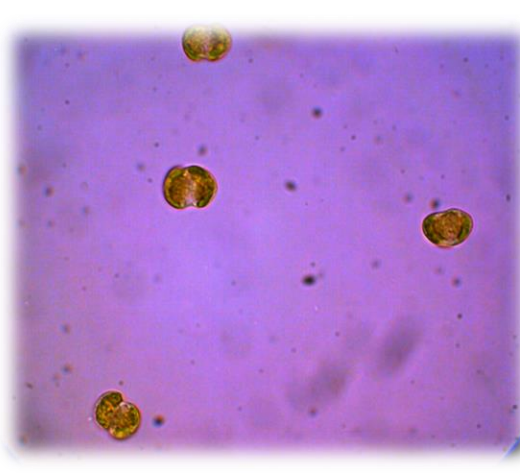
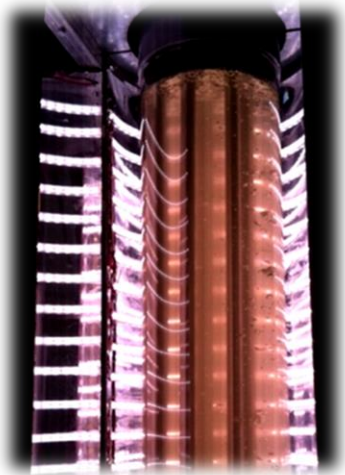
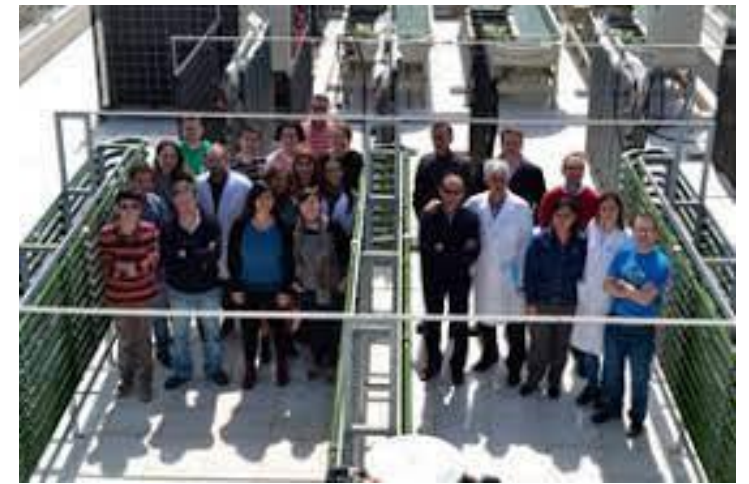
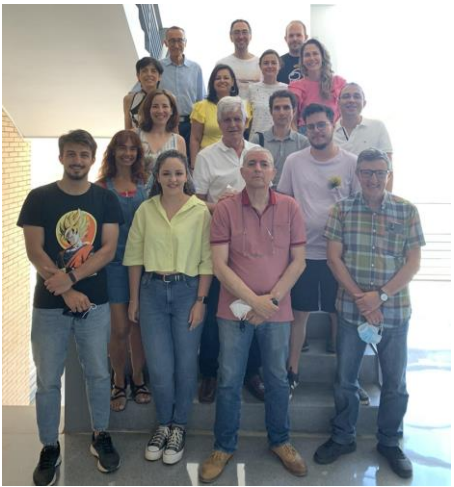


# Marine Microalgae Biotechnology (Biotechnological potential of marine dinoflagellates)



# Marine Microalgae Biotechnology Research Group (BIO173)



- Chemical Engineering Department
- Engineering treatment of bioprocesses

Members: **21**

Ph.D.: 16

Master: 5



Since late 1980s

>410 Papers in indexed Journals

44 Books/book chapters

> 440 Congress communications

> 180 Thesis/Master Thesis

58 Contracts (private and public organizations)

100 Research Projects

16 Patents

- L facilities
- C osed ph
- T harvesti
- E fractions
- r



Emilio Molina Grima



Asterio Sánchez Mirón



Francisco García Camacho

- roalgal biomass
- and downstream
- ication of biom

# 1. Overview

## OUTLINE

### 1. Overview

#### 2. Objectives

#### 3. Strategies

##### 3.1 Improvement of culture media

##### 3.2 Modelling the growth of dinoflagellates with ANNS

##### 3.3 Optimization of culture conditions

##### 3.4 Recycling recourses

##### 3.5 Scale-up of the bioprocess

#### 4. Conclusions

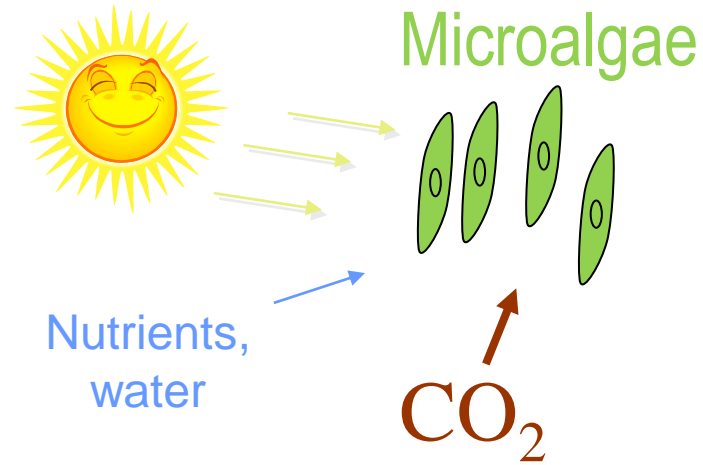
1.-Camacho, F. G., Rodríguez, J. G., Mirón, A. S., García, M. C., Belarbi, E. H., Chisti, Y., & Grima, E. M. (2007). Biotechnological significance of toxic marine dinoflagellates. *Biotechnology advances*, 25(2), 176-194.

2.-Gallardo-Rodríguez, J., Sánchez-Mirón, A., García-Camacho, F., **López-Rosales, L.**, Chisti, Y., & Molina-Grima, E. (2012). Bioactives from microalgal dinoflagellates. *Biotechnology advances*, 30(6), 1673-1684.

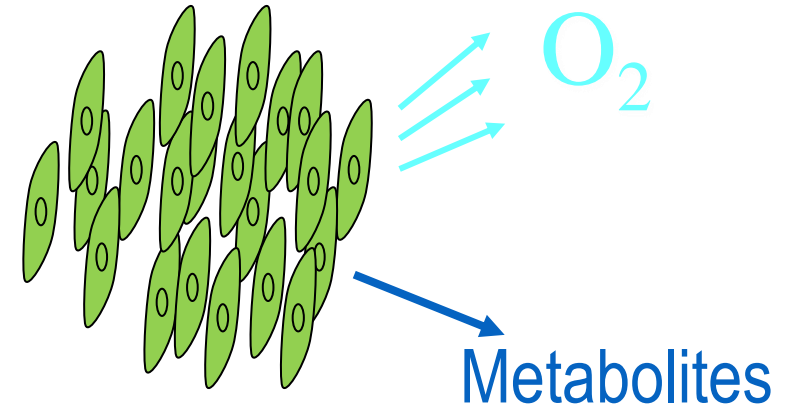
3.-García-Camacho, F., Sánchez-Mirón, A., Gallardo-Rodríguez, J., **López-Rosales, L.**, Chisti, Y., & Molina-Grima, E. (2014). Culture of microalgal dinoflagellates. In *Seafood and Freshwater Toxins: Pharmacology, Physiology, and Detection* (pp. 551-566). CRC Press Boca Raton.

# Marine Dinoflagellates

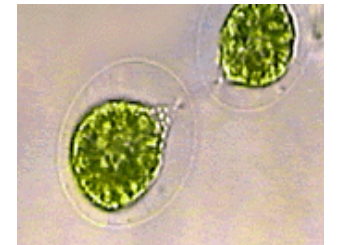
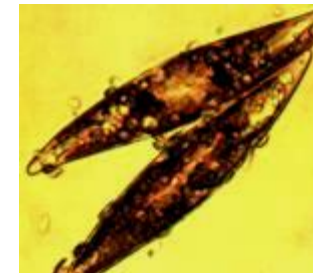
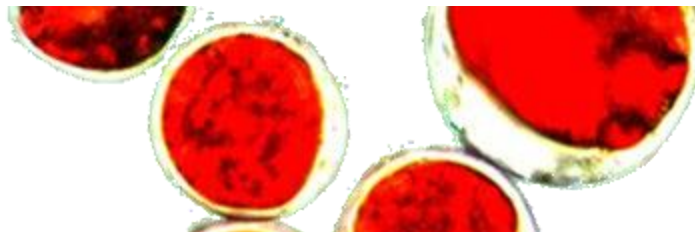
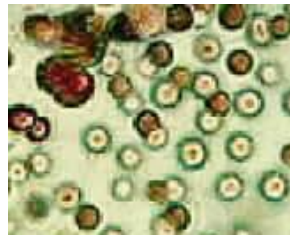
## Photoautotrophic unicellular microorganisms



## Microalgae



- High duplication speed

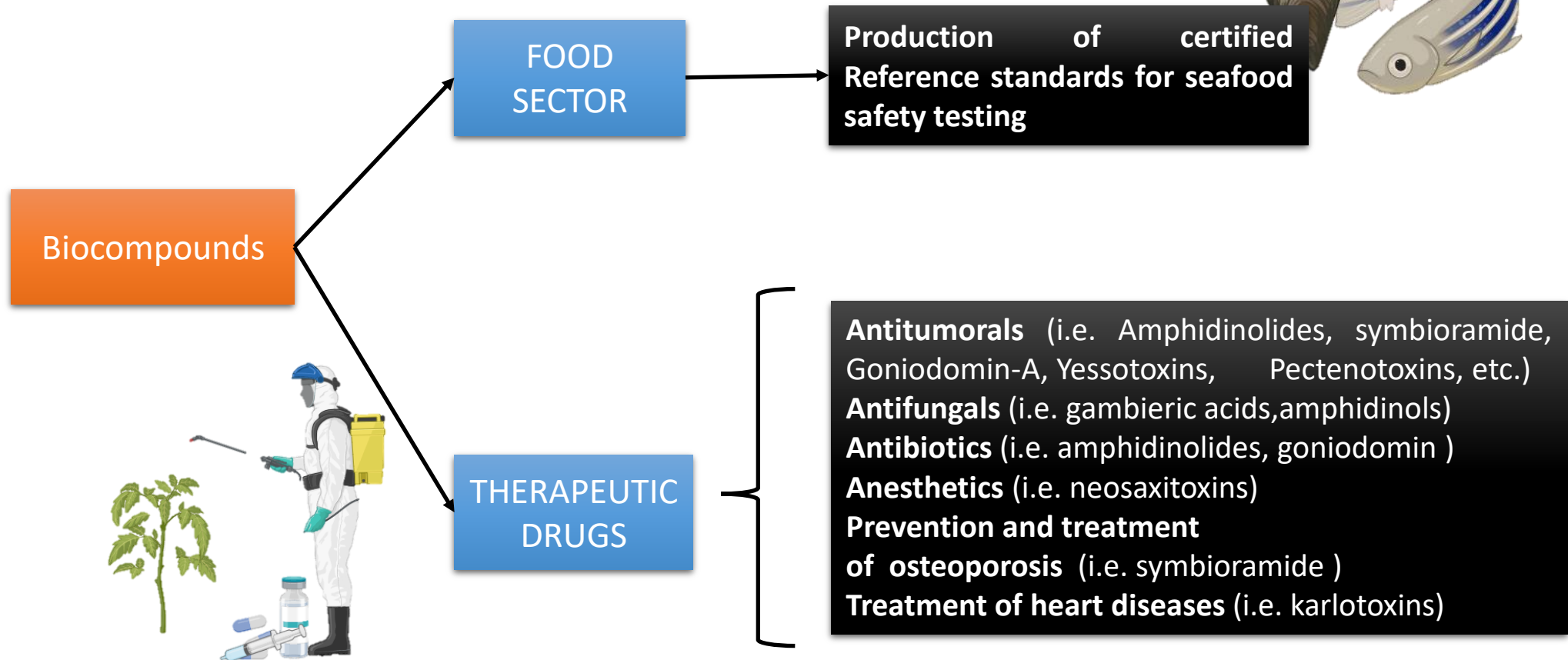


### OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

# Marine Dinoflagellates

- Microalgae producing **bioactive substances**



## OUTLINE

### 1. Overview

2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

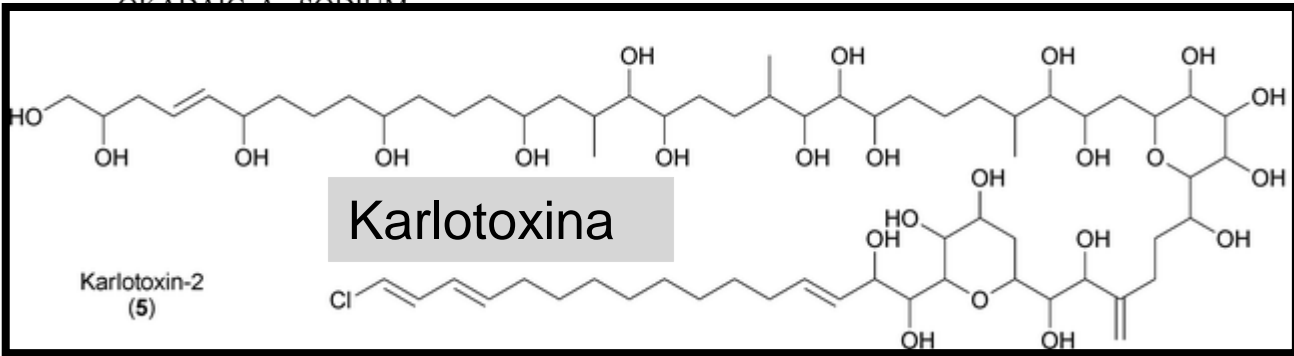
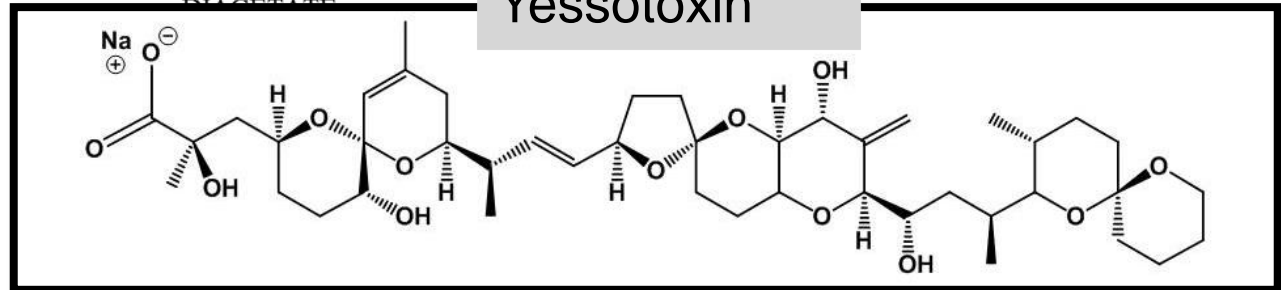


# Prices of commercial biotoxins from dinoflagellates

- **High structural complexity (no chemical synthesis)** from 2006 catalogues. The prices are calculated for 1 mg of toxin. More details about toxin characteristics, purity and extraction methods can be found in the corresponding references.
- **Large scale cultivation** -producing species indicated by suppliers were *Prorocentrum concavum*, *Ptychodiscus caribaeorum*, *Symyxium caribaeorum*, *Gambierdiscus toxicus*, *Palythoa caribaeorum*, y *Protogonyaulax sp*

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

	BIOTOXINAS	SGA	ALEX	LKT	VWR	WK*	LD	GEN	SL	PKC	TK
 <p>Karlotoxina</p> <p>Karlotoxin-2 (5)</p>	OKADAOL	5530€	-	-	3089€	-	4420€	-	1849€	676€	-
	SAXITOXIN, DIALGATELATE	13	-	-	-	-	-	-	3870€	676€	-
 <p>Yessotoxin</p> <p>DINOPHYSISTOXIN-1</p> <p>PECTENOTOXIN-6</p>	DINOPHYSISTOXIN-1	-	-	-	-	2125€	-	-	-	-	-
	PECTENOTOXIN-6	-	-	-	-	2125€	-	-	-	-	-

Okadaic acid

5980€/mg!

?

SGA; Sigma-Aldrich [www.sigmaaldrich.com](http://www.sigmaaldrich.com) GEN; Gentaaur Molecular Products [www.gentaaur.com](http://www.gentaaur.com) LD; The lab Depot [www.labdepotinc.com](http://www.labdepotinc.com) SL; ScienceLab.com [www.sciencelab.com](http://www.sciencelab.com) PKC; PKC pharmaceuticals Inc [www.lclabs.com](http://www.lclabs.com) TK; Tocris [www.tocris.com](http://www.tocris.com) BIO; WK; Wako Pure Chemical Industries Ltd [www.wakochem.com](http://www.wakochem.com) ALEX; Alexis [www.alexis-corp.com](http://www.alexis-corp.com) VWR; VWR International [www.vwrsp.com](http://www.vwrsp.com)

(\*) Los okadaicos de Wako proceden de la esponja marina *Alichondria okadae*

Gallardo-Rodríguez, J. et al. (2012). Bioactives from microalgal dinoflagellates. *Biotechnology Advances*. 30, 1673-1684.

# Drawbacks in the culture of marine dinoflagellates

- Very low bioactive titers ( $\mu\text{g L}^{-1}$ )
- Low cell densities ( $\text{mg L}^{-1}$  or  $10^{4-5}$  cell  $\text{ml}^{-1}$ )
- **Shear-sensitive species**
- Low growth rates
- Complex metabolism
- Nutritional requirements

## OUTLINE

### 1. Overview

#### 2. Objectives

#### 3. Strategies

##### 3.1 Improvement of culture media

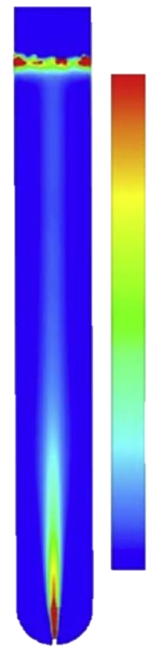
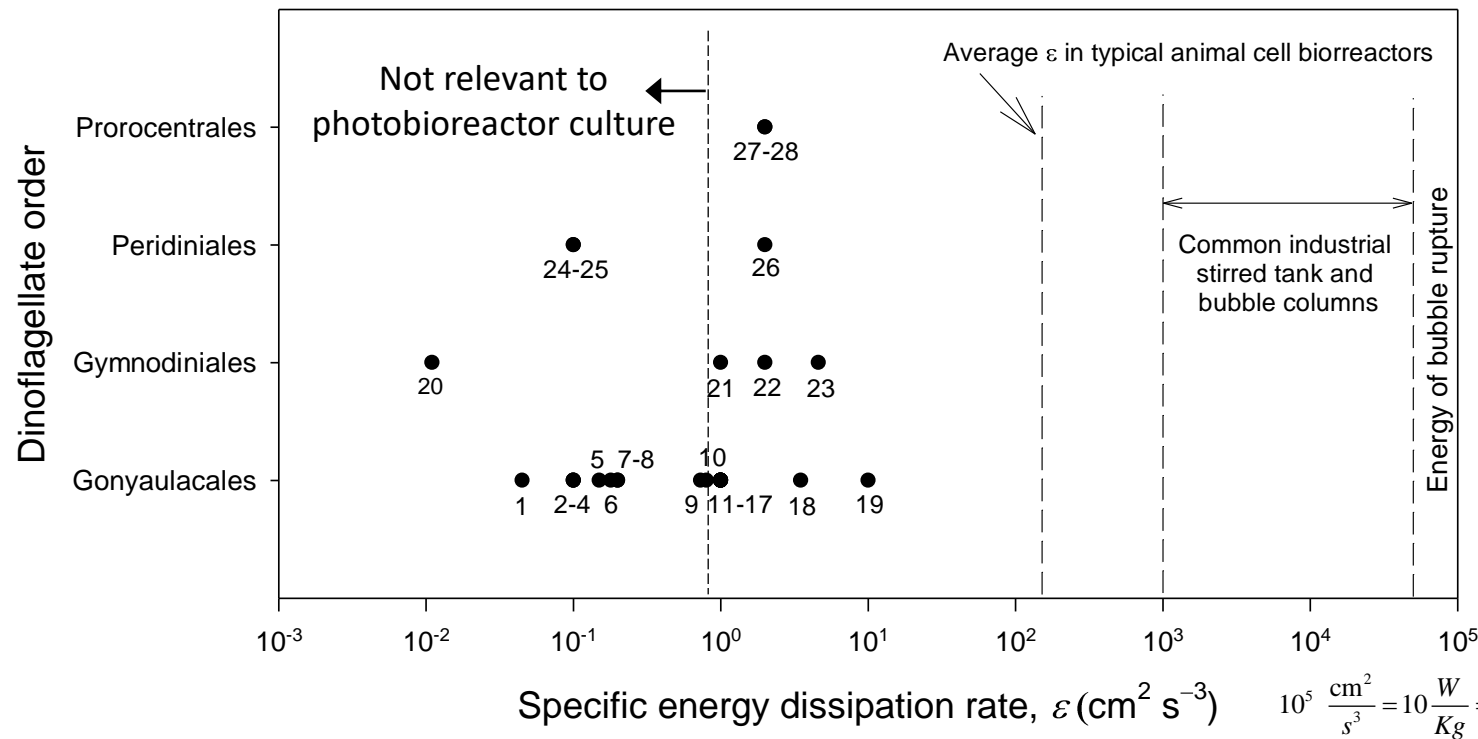
##### 3.2 Modelling the growth of dinoflagellates with ANNS

##### 3.3 Optimization of culture conditions

##### 3.4 Recycling recourses

##### 3.5 Scale-up of the bioprocess

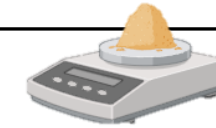
#### 4. Conclusions





# Amounts of product needed in a pharmaceutical application

- Discovery of potential drug candidate structure(s) by screening compound/extract libraries.
- Production for preclinical phases



**From several grams to hundreds of grams!**

- Production for clinical phases

**Multi-kilogram quantities**



**Major hurdle in the development of drugs from marine dinoflagellates**

**To ensure supply**



## OUTLINE

### 1. Overview

2. Objectives

3. Strategies

3.1 Improvement of culture media

3.2 Modelling the growth of dinoflagellates with ANNS

3.3 Optimization of culture conditions

3.4 Recycling recourses

3.5 Scale-up of the bioprocess

4. Conclusions

# 2. OBJECTIVES

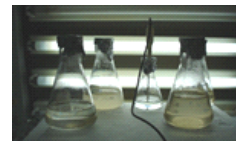
Conventional stirred tanks



Flat panel PBRs



Tubular PBRs



Shake flasks

Bubble columns



Tubular photobioreactors



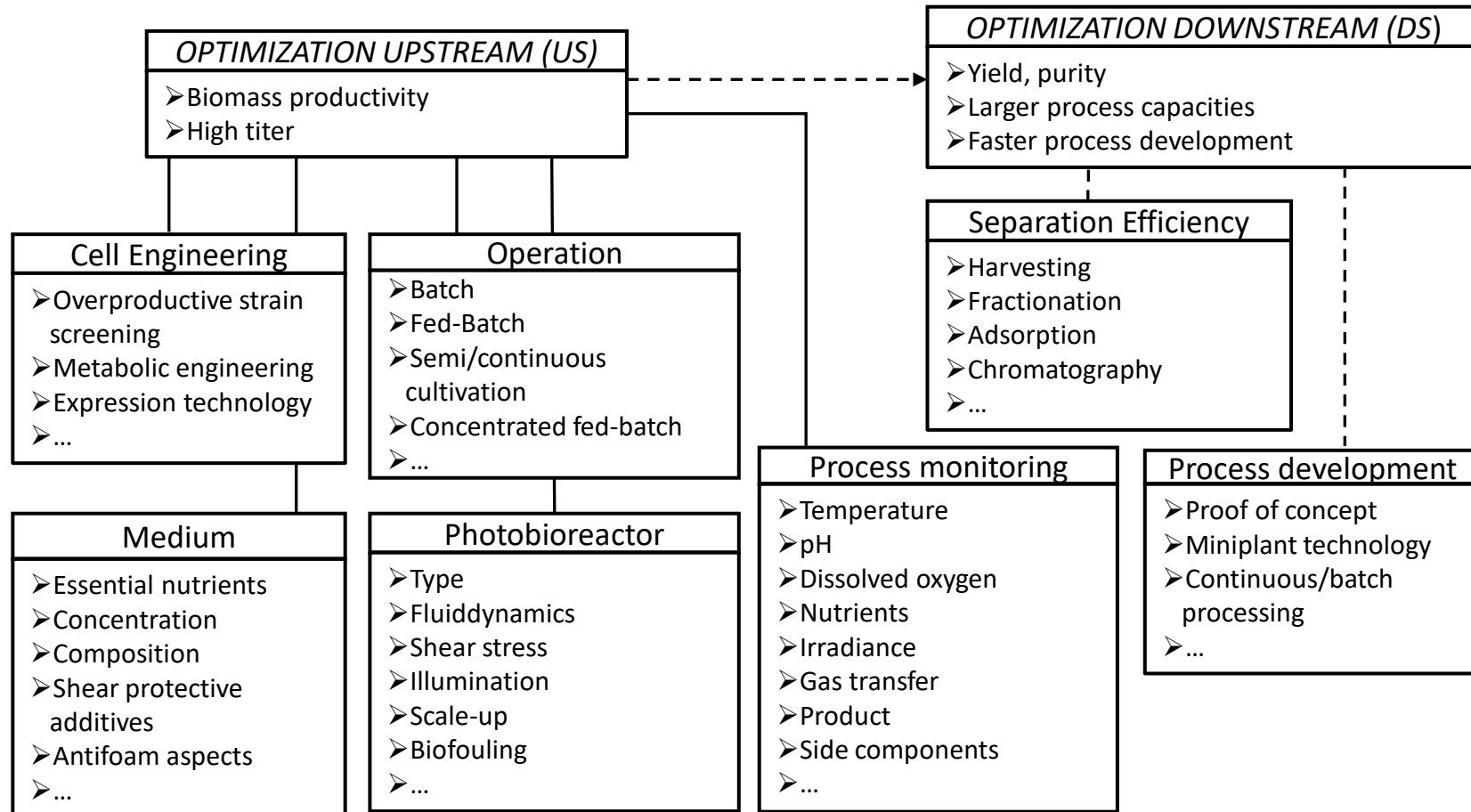
Volume	0.1–1	1–20	10–10 <sup>2</sup>	50–500	>10 <sup>3</sup>
Biomass	1–10 g L <sup>-1</sup>	0.01–0.3 g L <sup>-1</sup>		1–10 Kg m <sup>-3</sup>	1–300 g m <sup>-3</sup>

Photobioreactors typically used for culturing **non-dinoflagellate microalgae** at different scales of operation

## OUTLINE

1. Overview
2. **Objectives**
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

# How can bioactives from marine dinoflagellates become in drugs?



## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



# How can bioactives from marine dinoflagellates become in drugs?

1. Implementation of strategies for improving the **culture medium formulation**.
2. Analysis of **nutrient interactions** and evaluation of their **relative impact** on cell growth.
3. Study of **sensitivity** of dinoflagellate microalgae to turbulence developed in PBRs (energy dissipation rate, *EDR*).
4. Optimization of engineering factors influencing to **hydrodynamics** of bubble column PBRs for shear-sensitivity microalgae.
5. **Pilot-plant scale-up** of the culture in pneumatically agitated PBRs (indoor /outdoor conditions).

## OUTLINE

1. Overview
2. **Objectives**
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

# 3. Strategies

## 3.1. Improvement of culture media

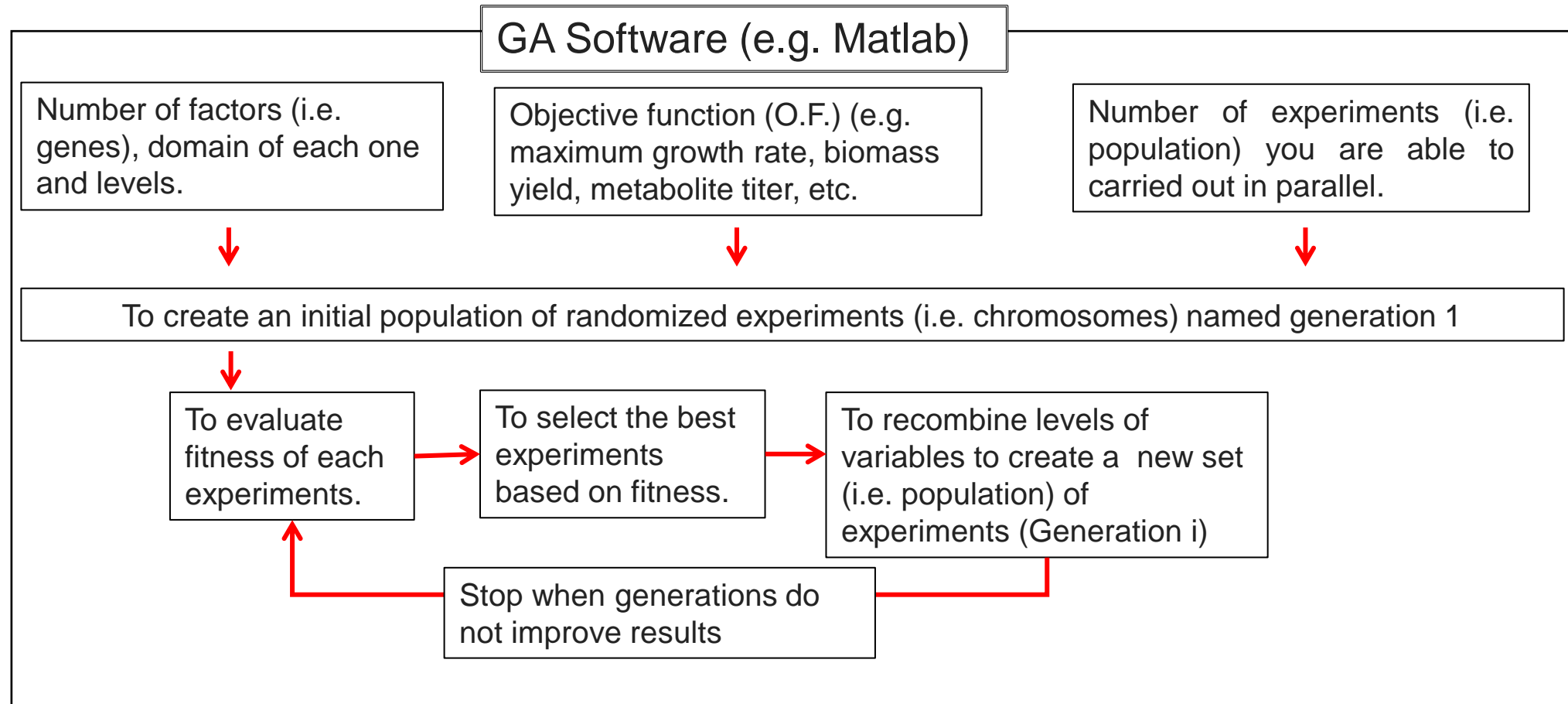
### OUTLINE

1. Overview
2. Objectives
3. Strategies
- 3.1 Improvement of culture media**
- 3.2 Modelling the growth of dinoflagellates with ANNS
- 3.3 Optimization of culture conditions
- 3.4 Recycling recourses
- 3.5 Scale-up of the bioprocess
4. Conclusions

**López-Rosales, L.,** García-Camacho, F., Sánchez-Mirón, A., & Chisti, Y. (2015). An optimal culture medium for growing *Karlodinium veneficum*: progress towards a microalgal dinoflagellate-based bioprocess. *Algal Research*, 10, 177-182.

# Improvement of culture media

✓ **Genetic algorithms (GA).** A class of stochastic search strategies inspired by the process of natural selection. GA are commonly used to **optimization** and search problems by relying on bio-inspired operators such as mutation, crossover and selection



## OUTLINE

1. Overview
2. Objectives
3. Strategies

### 3.1 Improvement of culture media

3.2 Modelling the growth of dinoflagellates with ANNS

3.3 Optimization of culture conditions

3.4 Recycling recourses

3.5 Scale-up of the bioprocess

4. Conclusions

# Improvement of culture media (*Karlodinium veneficum*)

## OUTLINE

1. Overview
2. Objectives
3. Strategies

### 3.1 Improvement of culture media

### 3.2 Modelling the growth of dinoflagellates with ANNS

### 3.3 Optimization of culture conditions

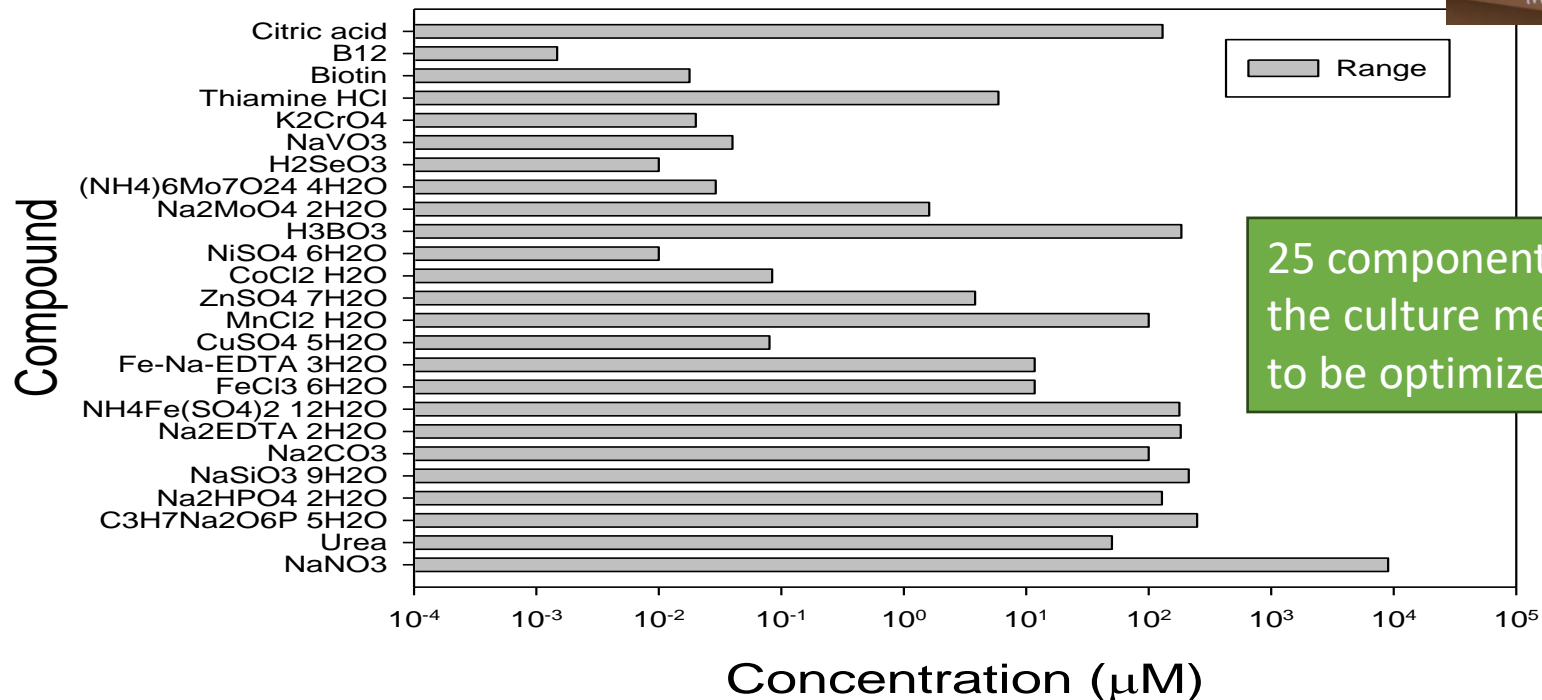
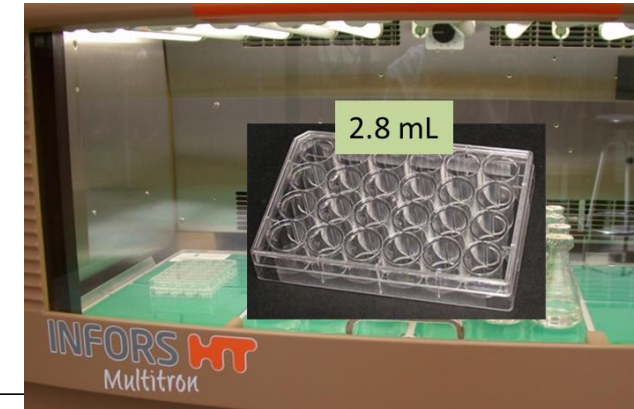
### 3.3 New aspects on the sensitivity of dinoflagellates to hydrodynamic stress

### 3.4 Recycling recourses

### 3.5 Scale-up of the bioprocess

## 4. Conclusions

**Photobioreactors:** Polystyrene multiwell plates (24-wells; Corning®) placed in incubator. **Volume:** 2.8 mL. **Initial concentration:**  $3 \times 10^4$  cells·mL<sup>-1</sup>. **Irradiance:** 200  $\mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$  (12:12h light–dark regimen) from daylight fluorescent lights. **Temperature:**  $18 \pm 1$  ° The optimization experiments were in triplicate.



25 components of the culture medium to be optimize

# Improvement of culture media (*Karlodinium veneficum*)

## OUTLINE

1. Overview
2. Objectives
3. Strategies

### 3.1 Improvement of culture media

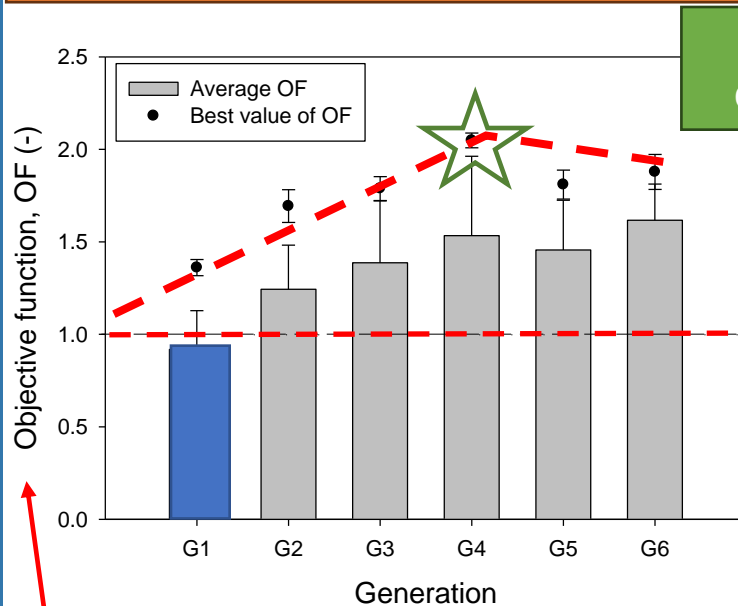
### 3.2 Modelling the growth of dinoflagellates with ANNS

### 3.3 Optimization of culture conditions

### 3.4 Recycling recourses

### 3.5 Scale-up of the bioprocess

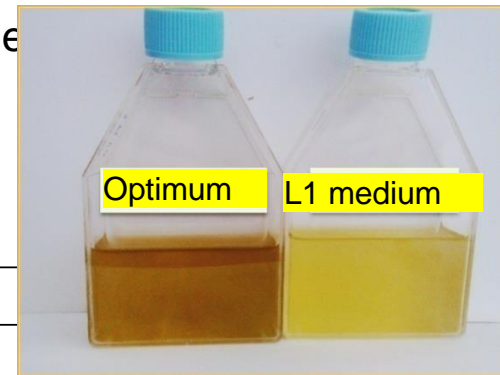
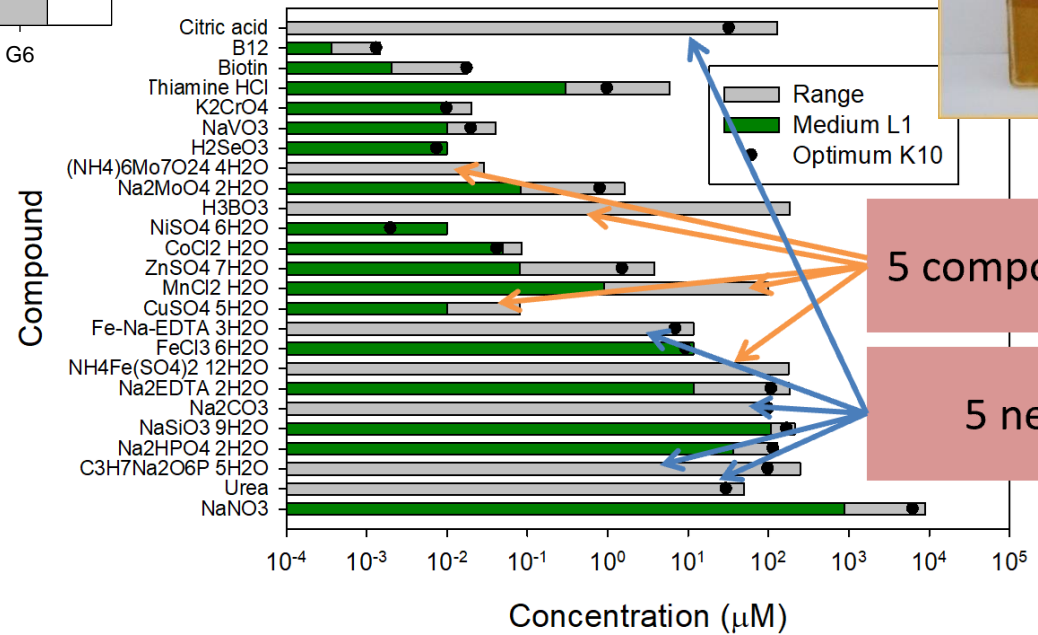
## 4. Conclusions



Optimum composition

Maximum number of individuals (i.e. experiments) in a single generation = 70 cultures.  
420 experiments, each with a different media composition

O.F = biomass productivity relative to the control culture grown in the L1 medium



5 compounds were ruled out

5 new components



# Improvement of culture media (*Karlodinium veneficum*)

## OUTLINE

1. Overview
2. Objectives
3. Strategies

### 3.1 Improvement of culture media

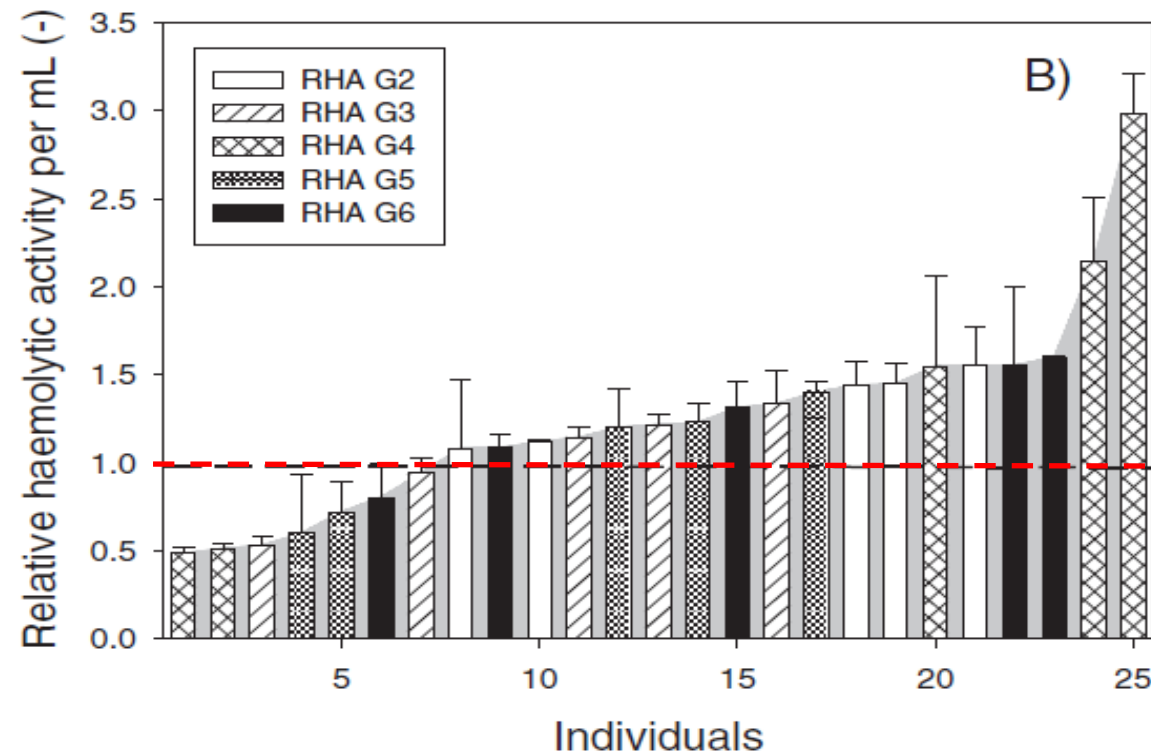
3.2 Modelling the growth of dinoflagellates with ANNS

3.3 Optimization of culture conditions

3.4 Recycling recourses

3.5 Scale-up of the bioprocess

4. Conclusions



Most of them exceeded the control

**Fig..** Relative hemolytic activity (RHA) per per milliliter of culture supernatan for the five best media formulations of each generation. The dashed line indicates the RHA for the L1 control culture.

# 3. Strategies

## 3.2. Modelling the growth of dinoflagellates with artificial neural networks

### OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS**
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

García-Camacho, F., **López-Rosales, L.**, Sánchez-Mirón, A., Belarbi, E. H., Chisti, Y., & Molina-Grima, E. (2016). Artificial neural network modeling for predicting the growth of the microalga *Karlodinium veneficum*. *Algal Research*, 14, 58-64.

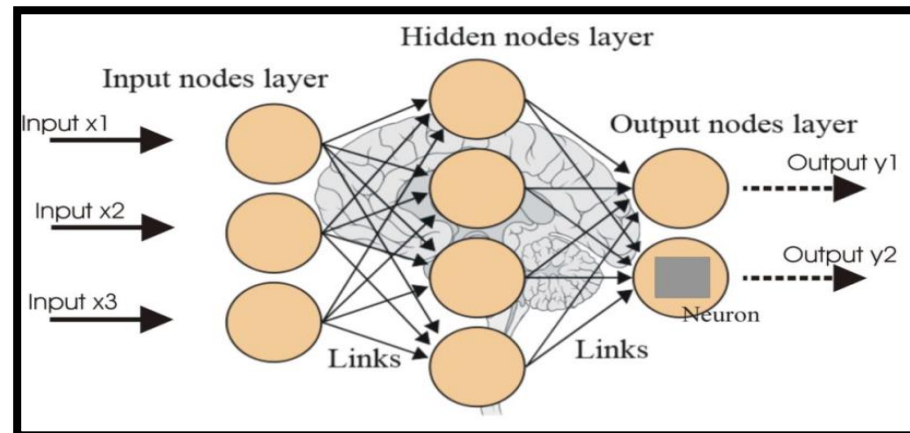
**López-Rosales, L.**, Gallardo-Rodríguez, J. J., Sánchez-Mirón, A., Contreras-Gómez, A., García-Camacho, F., & Molina-Grima, E. (2013). Modelling of multi-nutrient interactions in growth of the dinoflagellate microalga *Protoceratium reticulatum* using artificial neural networks. *Bioresource technology*, 146, 682-688.

# What does Artificial Neural Network (ANN) mean?

- It is a **computational model** based on the structure and functions of **biological neural networks**.
- ANNs solves problems that would prove impossible or difficult by human or statistical standards
- ANN has **self-learning capabilities** that enable it produce better results as more data becomes available.
- The **structure** of the ANN is function of Information that flows through it (i.e. **typology of the inputs and output**)
- ANNs are considered **nonlinear statistical data modeling tools** where the **complex** relationships between inputs and outputs are modeled or patterns are found.

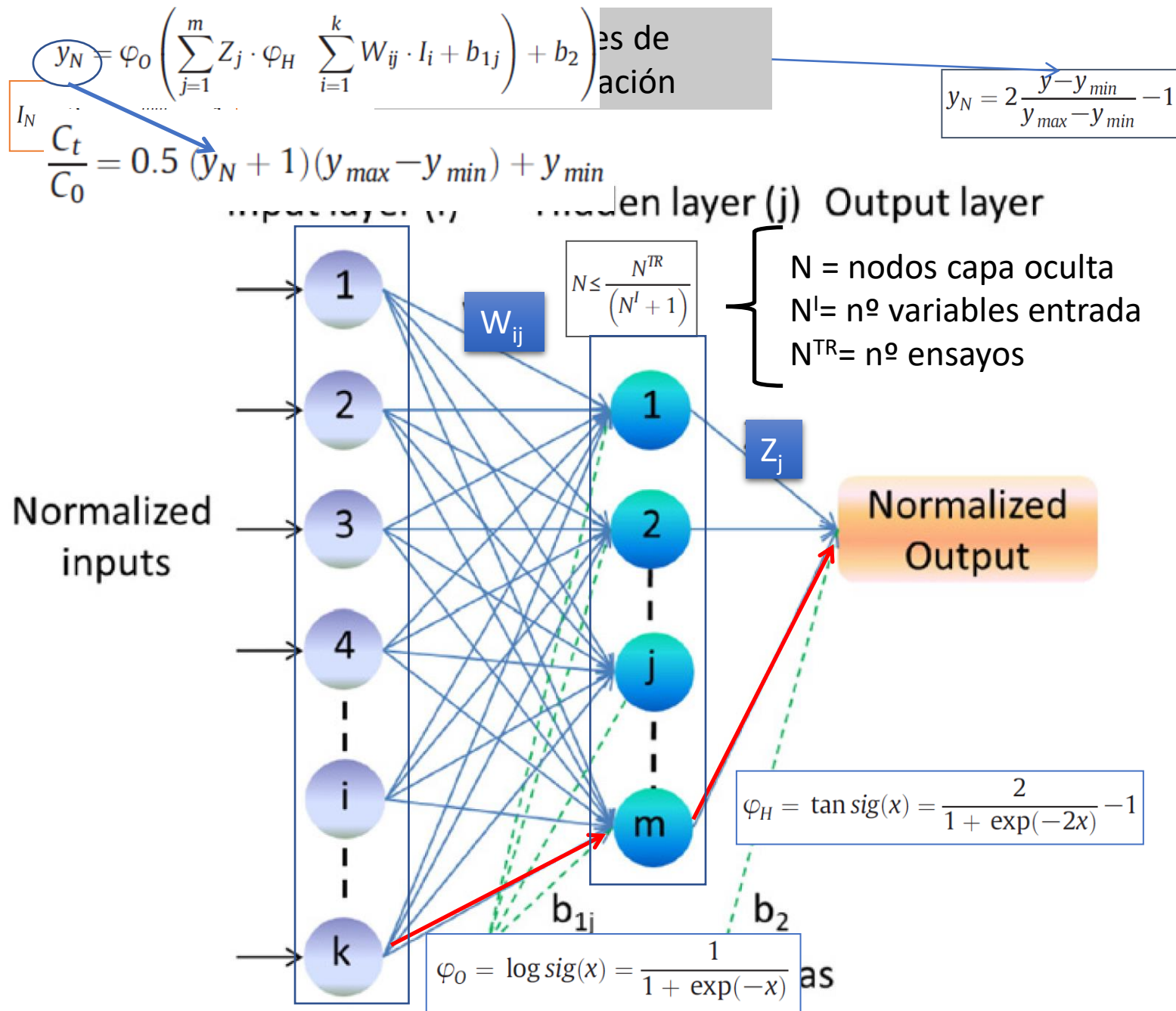
## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS**
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS**
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



# ANN modeling for predicting the growth of the microalga *Karlodinium veneficum*

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

**ANN input variable.** a matrix (k x m) whose columns were vectors with the concentrations ( $\mu\text{M}$ ) of 25 components selected, initial cell concentration ( $C_0$ ) and culture time (t).

**Target variable:** vector with  $C_t/C_0$ .

## Implementation of the ANN model formulation in Matlab

Generation	1	1	1	1	1	5	5	5	5	5	5	5	5	5
t, days	0	0	6	6	6	3	3	3	3	3	6	6	6	6
[Co], cells/mL	33000	33	67	33000	67	67	33	67	31000	33	31000	67	29000	32000
NaNO <sub>3</sub>	0	0	2700	5400	3600	900	8100	2700	882	1800	8100	3600	4500	6300
Urea	20	20	30	30	10	20	30	20	0	20	10	30	50	40
Glycerophosphate	99.2	148.8	74.4	24.8	49.6	74.4	173.6	74.4	0	173.6	173.6	173.6	99.2	74.4
PO <sub>4</sub> <sup>-3</sup>	64	115.2	25.6	12.8	12.8	89.6	115.2	128	36.2	76.8	128	12.8	102.4	115.2
SiO <sub>4</sub> <sup>-2</sup>	42.4	42.4	169.6	42.4	169.6	42.4	84.8	42.4	106	127.2	127.2	169.6	42.4	84.8
CO <sub>2</sub> <sup>-2</sup>	60	20	20	80	20	100	20	100	0	60	60	20	60	100
Na <sub>2</sub> EDTA·2H <sub>2</sub> O	36.46	109.38	182.3	0	145.84	109.38	0	0	11.7	145.84	145.84	36.46	36.46	145.84
Fe(NH <sub>4</sub> ) <sub>2</sub> (SO <sub>4</sub> ) <sub>2</sub> ·6H <sub>2</sub> O	0	89	178	89	44.5	0	133.5	44.5	0	44.5	44.5	0	0	0
FeCl <sub>3</sub> ·6H <sub>2</sub> O	4.68	11.7	7.02	2.34	9.36	9.36	11.7	0	11.7	0	0	7.02	4.68	9.36
Fe-Na-EDTA·3H <sub>2</sub> O	7.02	2.34	2.34	7.02	2.34	7.02	4.68	9.36	0	7.02	7.02	7.02	7.02	4.68
CuSO <sub>4</sub> ·5H <sub>2</sub> O	0.06	0.04	0.04	0.06	0.06	0	0.04	0.04	0.01	0.04	0.04	0.04	0.06	0.04
MnCl <sub>2</sub> ·H <sub>2</sub> O	20	60	80	70	70	0	90	30	0.91	30	30	0	20	50
ZnSO <sub>4</sub> ·7H <sub>2</sub> O	2.674	3.056	1.91	3.82	2.292	1.528	0.382	3.438	0.08	2.292	2.292	1.528	2.674	2.292
CoCl <sub>2</sub> ·6H <sub>2</sub> O	0.056	0.056	0.042	0.07	0.056	0.042	0.056	0.028	0.05	0.014	0.042	0.014	0.056	0.014
NiSO <sub>4</sub> ·6H <sub>2</sub> O	0.008	0.002	0.004	0.01	0.006	0.008	0.002	0.01	0.01	0	0.002	0.006	0.008	0.004
H <sub>3</sub> BO <sub>3</sub>	74	74	111	148	37	111	74	185	0	148	0	111	74	111
Na <sub>2</sub> MoO <sub>4</sub> ·2H <sub>2</sub> O	1.072	0.804	0	1.34	0	1.34	0.268	1.072	0.0822	1.072	0.804	0.536	1.072	0.536
(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub> ·4H <sub>2</sub> O	0.00728	0.01456	0.02184	0.01456	0	0.02912	0.02912	0.02184	0	0.02184	0	0.00728	0.00728	0.02184
H <sub>2</sub> SeO <sub>3</sub>	0.005	0.01	0.005	0.01	0.01	0.005	0	0	0.01	0.005	0.01	0.01	0.005	0.0025
Na <sub>3</sub> VO <sub>4</sub>	0.02	0.03	0.01	0.02	0.03	0.03	0	0.02	0.01	0.03	0.02	0.03	0.02	0.03
K <sub>2</sub> CrO <sub>4</sub>	0	0.01	0.02	0.015	0.02	0.02	0	0.01	0.01	0	0	0.01	0	0
Thiamine·HCl	0	2.964	0	4.94	4.94	2.964	0.988	4.94	0.296	3.952	0.988	0	0	2.964
Biotin	0.00712	0.01246	0.01424	0.01424	0.00356	0.01068	0.00356	0.01424	0.002045	0.01424	0.00356	0.01424	0.00712	0.00356
Vitamin B <sub>12</sub>	0.000296	0.000296	0	0.000296	0.000888	0.000296	0.001332	0.000888	0.000369	0.001184	0.000592	0.00074	0.000296	0.000888
Citric Acid·H <sub>2</sub> O	32.5	130	0	65	65	97.5	97.5	97.5	0	32.5	97.5	65	32.5	97.5

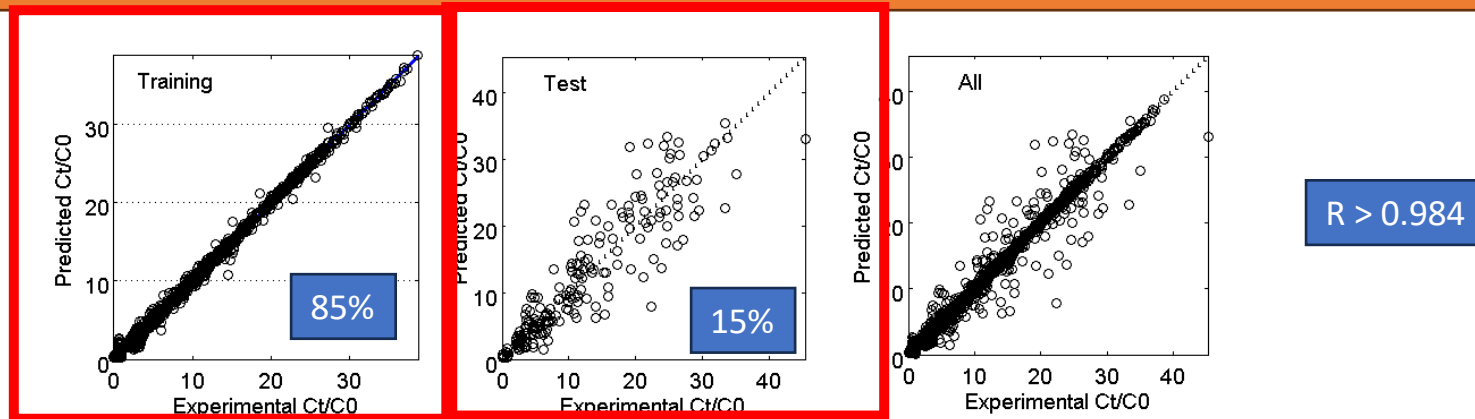
m = 1398

k = 27

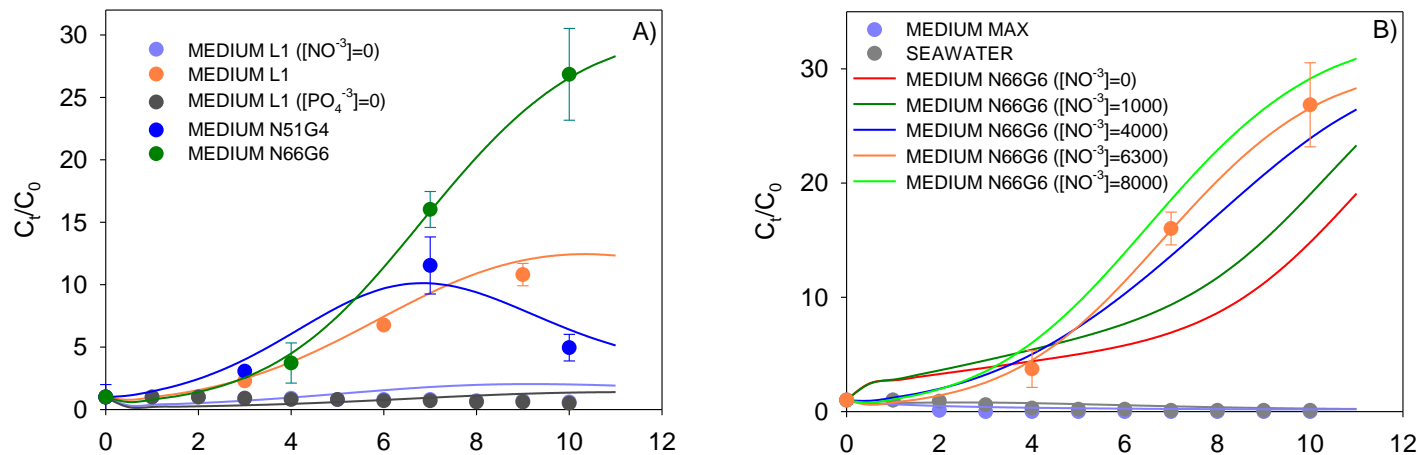
# ANN modeling for predicting the growth of the microalga *Karlodinium veneficum*

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS**
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



A parity plot comparison of the predicted and the measured (experimental) dimensionless cell concentrations for the training runs, the test runs and all the runs combined.

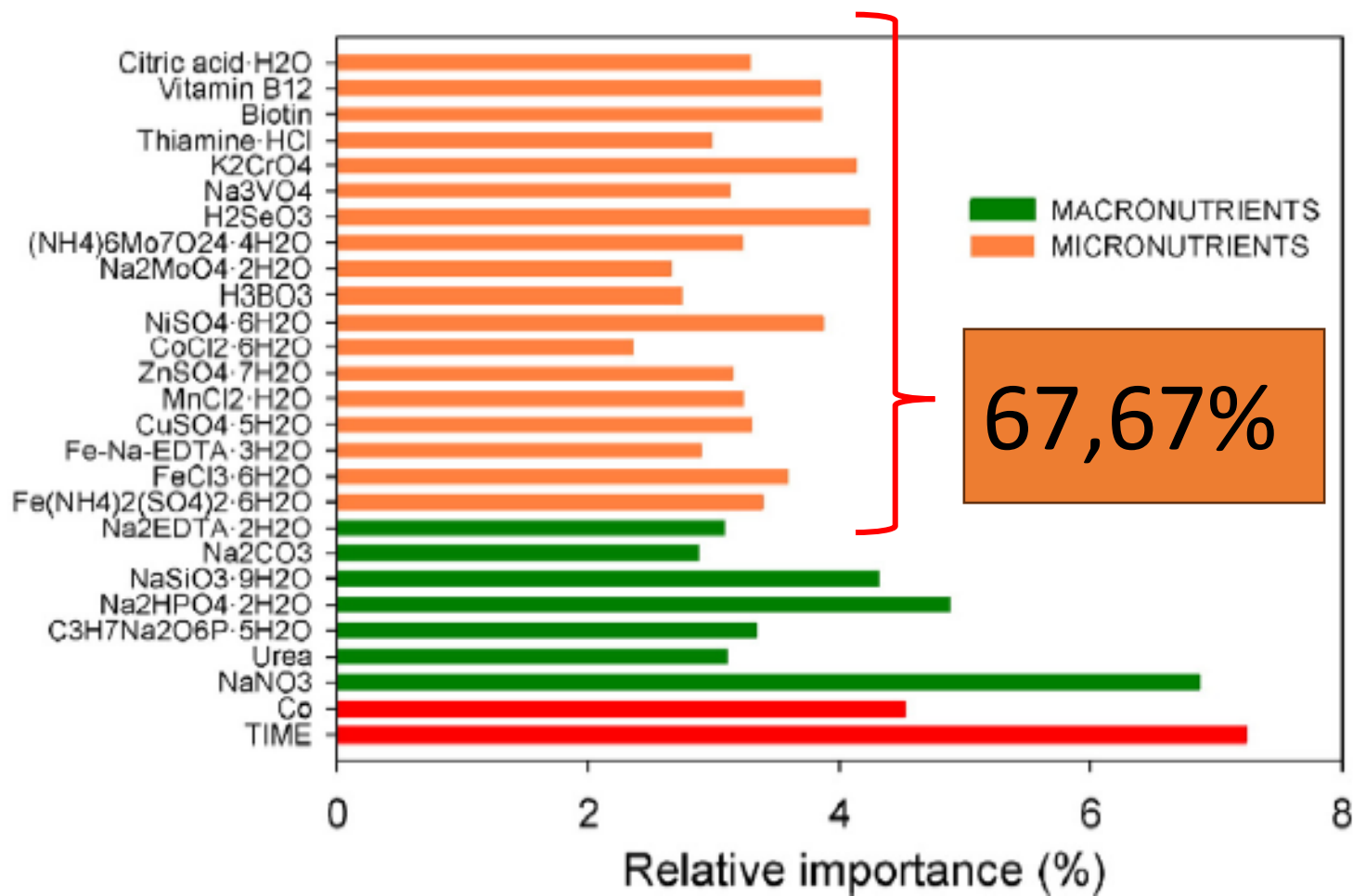


Examples of growth kinetics simulated by FBN model for different formulations of culture media. Solids circles represent experimental data. Predictions of the FBN model are shown as solid lines.

# ANN modeling for predicting the growth of the microalga *Karlodinium veneficum*

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS**
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



# 3. Strategies

## 3.3. Optimization of culture conditions

### OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions**
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

**López-Rosales, L.,** García-Camacho, F., Sánchez-Mirón, A., Contreras-Gómez, A., & Molina-Grima, E. (2015). An optimisation approach for culturing shear-sensitive dinoflagellate microalgae in bench-scale bubble column photobioreactors. *Bioresource technology*, 197, 375-382.

**López-Rosales, L.,** García-Camacho, F., Sánchez-Mirón, A., Contreras-Gómez, A., & Molina-Grima, E. (2017). Modeling shear-sensitive dinoflagellate microalgae growth in bubble column photobioreactors. *Bioresource technology*, 245, 250-257.



# An optimisation approach for culturing *K. veneficum* in bench-scale bubble column PBRs

✓ **Bank of bench-scale bubble column photobioreactors** (4.4 cm internal diameter) to find an optimal combination of values for the culture parameters gas flow rate, culture height, and nozzle sparger diameter

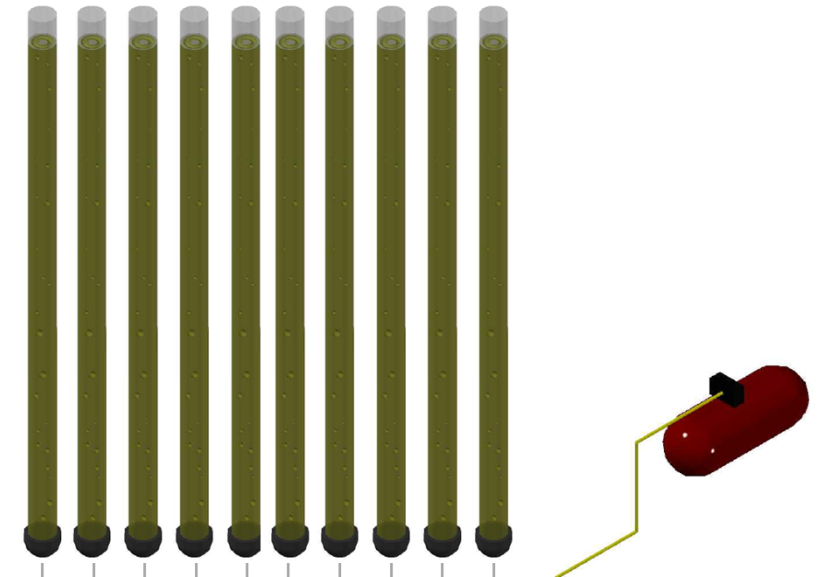
## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions**
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



Factor	Units	Range of variation	Step-width
Air flow rate ( $Q$ )	$\text{L min}^{-1}$	0.1–0.5	0.13 (4)
Culture height ( $H$ )	m	0.50–1.75	25 (6)
Nozzle diameter ( $d_0$ )	mm	1–2.5	0.5 (4)

The numbers in parentheses represent the number of possible levels for each factor. The product of these values gives the total number of possible experiments ( $4 \times 6 \times 4 = 96$ ).



# An optimisation approach for culturing *K. veneficum* in bench-scale bubble column PBRs

## OUTLINE

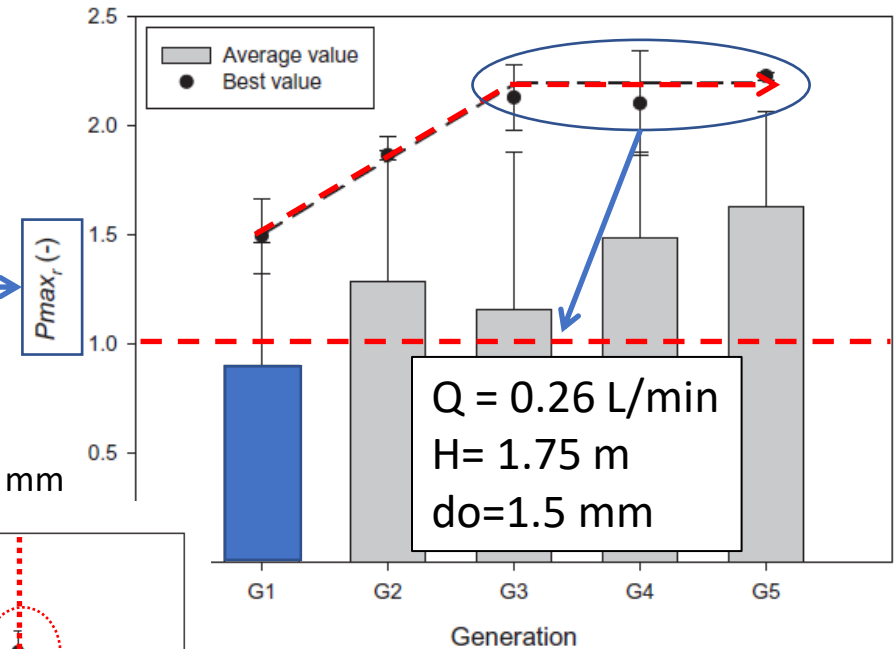
1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions**
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

Optimization technique:  
Genetic algorithm

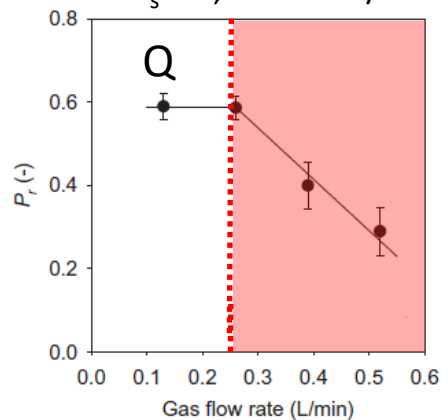
Number of individuals (i.e. experiments) in a single generation = 5 batch cultures.

25 experiments, each with a different combination of Q, H and  $d_o$ .

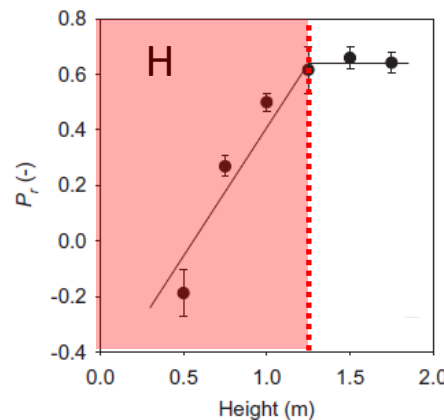
O.F. = maximum biomass productivity ( $P_{max,r}$ ) relative to the static control culture in T-Flask



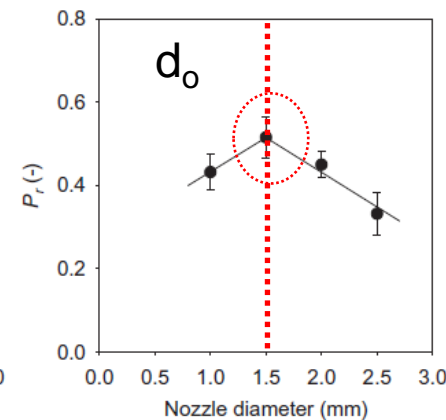
$Q < 0.26$  L/min  
or 0,137 vvm  
 $v_s < 2,85 \cdot 10^{-3}$  m/s



$H > 1.25$  m



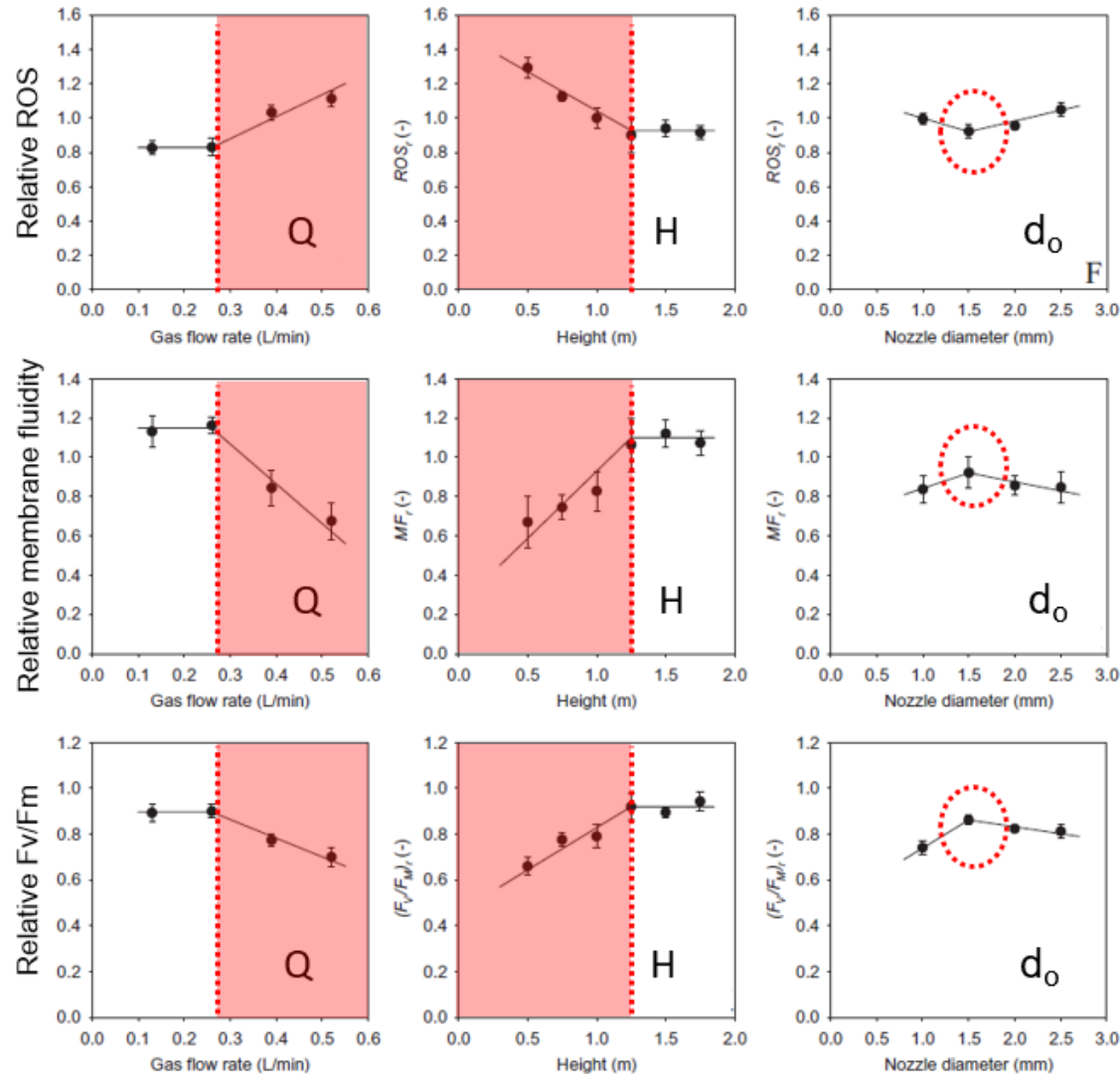
$d_o = 1.5$  mm



# An optimisation approach for culturing *K. veneficum* in bench-scale bubble column PBRs

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions**
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



ROS  $\uparrow$  MF  $\downarrow$  Fv/Fm  $\downarrow$

Damage by hydrodynamic stress

Work window

Q(Lpm)	H(m)	do(mm)
<0.26	>1.25	1.5

# 3. Strategies

## 3.4. Recycling recourses

### OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses**
  - 3.5 Scale-up of the bioprocess
4. Conclusions

**L. López-Rosales**, P. López-García , M.A. Benyachou , A. Molina-Miras , J.J. Gallardo-Rodríguez , M.C. Cerón-García , A. Sánchez Mirón , F. García-Camacho(2022). Treatment of secondary urban wastewater with a low ammonium-tolerant marine microalga using zeolite-based adsorption. Bioresource Technology 359

A Molina-Miras, **L. López-Rosales**, MC Cerón-García, A Sánchez-Mirón, **A Olivera-Gálvez**, F García-Camacho, Molina-Grima (2020). Acclimation of the microalga *Amphidinium carterae* to different nitrogen sources: potential application in the treatment of marine aquaculture effluents. Journal of applied phycology



# Recycling recourses: Use of urban wastewater

Journal of Applied Phycology

<https://doi.org/10.1007/s10811-020-02049-9>

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses**
  - 3.5 Scale-up of the bioprocess
4. Conclusions

## Acclimation of the microalga *Amphidinium carterae* to different nitrogen sources: potential application in the treatment of marine aquaculture effluents



A. Molina-Miras<sup>1</sup> · L. López-Rosales<sup>1</sup> · M. C. Cerón-García<sup>1</sup> · A. Sánchez-Mirón<sup>1</sup> · A. Olivera-Gálvez<sup>2</sup> · F. García-Camacho<sup>1</sup> · E. Molina-Grima<sup>1</sup>

The initial treatment of wastewater is necessary to lower the concentration of ammoniacal nitrogen.

$\text{NH}_4^+-\text{N}_{\text{UWW}} = 54 \text{ mg/L}$

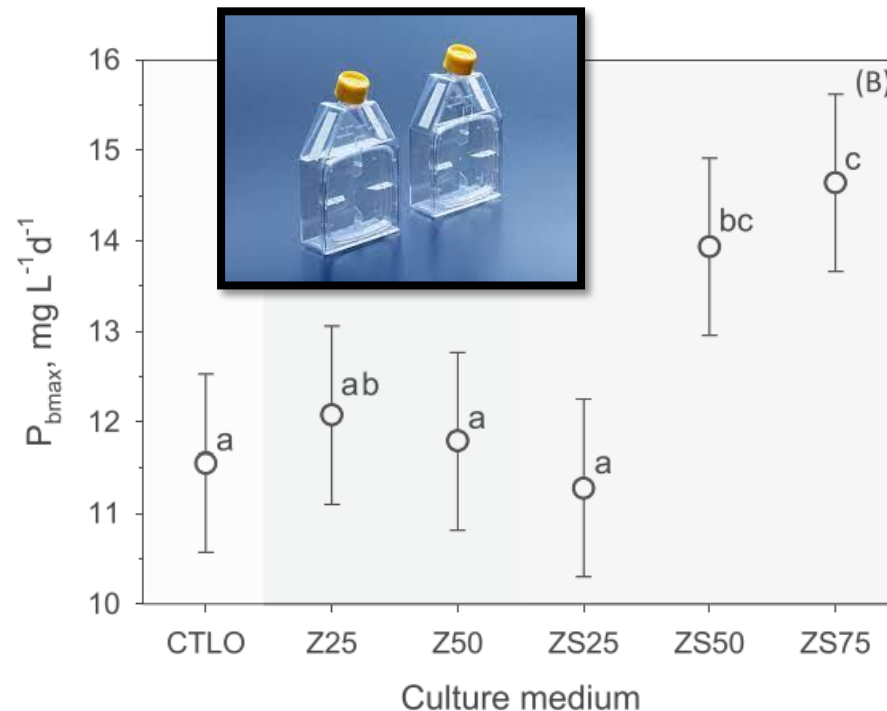
$\text{NH}_4^+-\text{N}_{\text{tox}} \approx 6 \text{ mg/L}$

# Recycling recourses: Use of urban wastewater

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses**
  - 3.5 Scale-up of the bioprocess
4. Conclusions

Characteristics	Culture media													UWWS	UWW
	CTLO	CTL1	CTL2	CTL3	CTL4	ZS25	Z25	ZS50	Z50	ZS75	Z75	ZS100	Z100		
UWW (%)	-	-	-	-	-	-	-	-	-	-	-	-	-	100	100
Zeolite-treated UWW (%)	-	-	100	50	-	25	25	50	50	75	75	100	100	-	-
Seawater (%)	100	100	-	50	-	75	75	50	50	25	25	-	-	-	-
Zeolite-treated SW (%)	-	-	-	-	100	-	-	-	-	-	-	-	-	-	-
Salinity Adjustment	No	No	Yes	Yes	No	Yes	No	Yes	No	Yes	No	Yes	No	Yes	No
Salinity (‰)	38.0	38.0	38.0	38.0	38.0	38.0	28.9	38.0	19.7	38.0	10.6	38.0	1.4	38.0	1.4
$[\text{NH}_4^+\text{-N}]_0$ , mg L <sup>-1</sup>	0.0	0.0	0.0	0.0	0	2.3	2.3	4.6	4.6	7.0	7.0	9.3	9.3	54.4	54.4
$[\text{NO}_3^-\text{-N}]_0$ , mg L <sup>-1</sup>	12.3	0.0	0.0	0.0	12.3	12.3	12.3	12.3	12.3	12.3	12.3	12.3	12.3	2.1	2.1
N <sub>TO</sub> , mg L <sup>-1</sup>	12.3	0.0	0.0	0.0	12.3	14.6	14.6	16.9	16.9	19.3	19.3	21.6	21.6	56.5	56.5
$[\text{PO}_4^{3-}\text{-P}]_0$ , mg L <sup>-1</sup>	5.4	0.0	0.0	0.0	5.4	5.4	5.4	5.4	5.4	5.4	5.4	5.4	5.4	0.8	0.8
$[\text{TOC}]_0$ , mg L <sup>-1</sup>	11.2	10.8	23.2	17	13.9	14.6	14.6	18	18	21.5	21.5	24.9	24.9	31.6	31.6

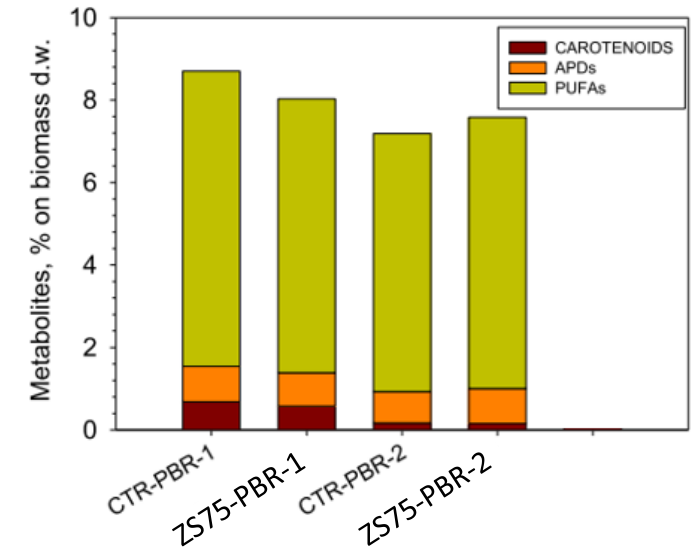
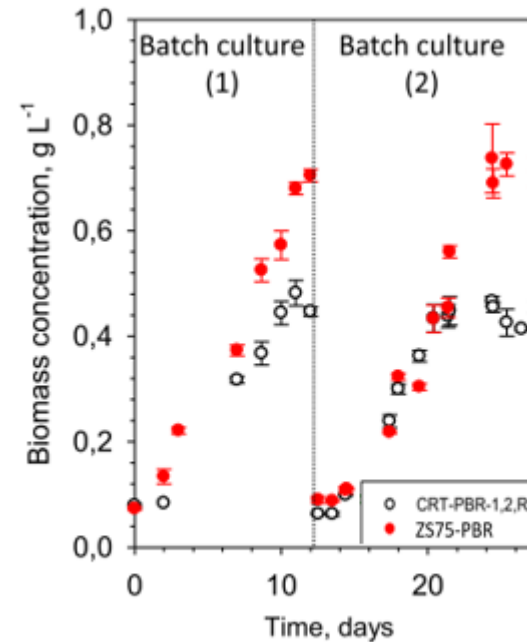
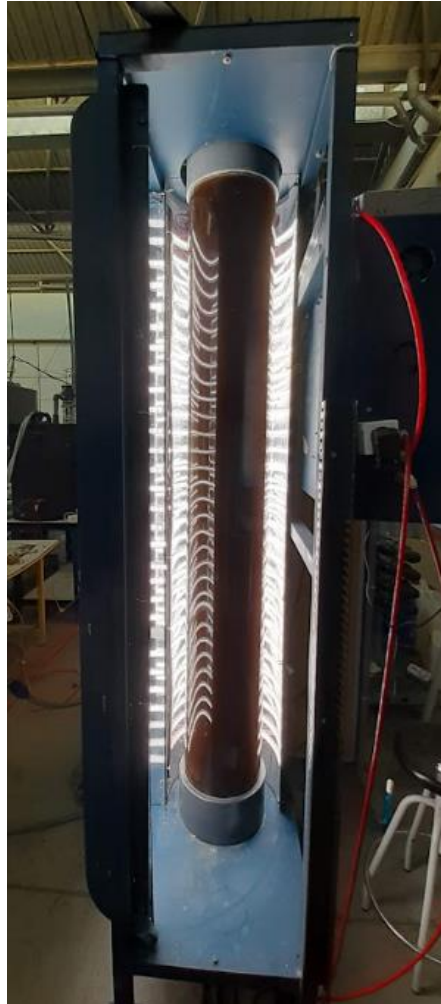


- No statistically significant differences between ZS50 and ZS75
- Without growth in the media that does not adjust the saline content
- The culture medium formulated with 75% urban wastewater treated with zeolites (7 mg/L NH<sub>4</sub><sup>+</sup>-N) adjusting the salinity obtained the best results in productivity of maximum biomass generated

# Recycling recourses: Use of urban wastewater

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses**
  - 3.5 Scale-up of the bioprocess
4. Conclusions

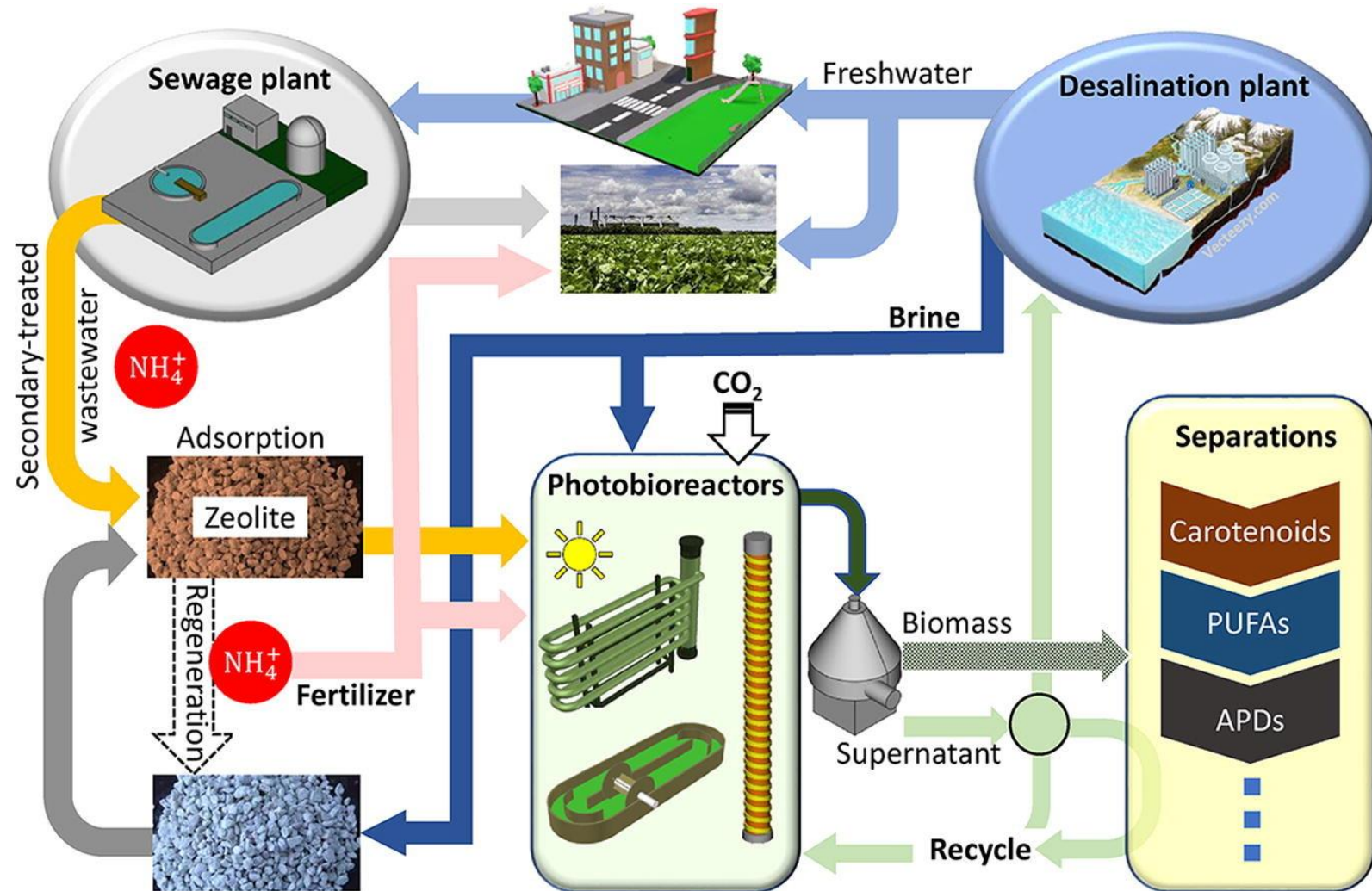


- Significant increase in biomass compared to the control culture
- No differences between the control culture and ZS75 in terms of dry weight percentage of biocompounds, but in terms of productivity

# Recycling recourses: Use of urban wastewater

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses**
  - 3.5 Scale-up of the bioprocess
4. Conclusions





# 3. Strategies

## 3.5. Scale-up of the bioprocess

### OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess**
4. Conclusions

**López-Rosales, L.,** García-Camacho, F., Sánchez-Mirón, A., Contreras-Gómez, A., & Molina-Grima, E. (2015). An optimisation approach for culturing shear-sensitive dinoflagellate microalgae in bench-scale bubble column photobioreactors. *Bioresource technology*, 197, 375-382.

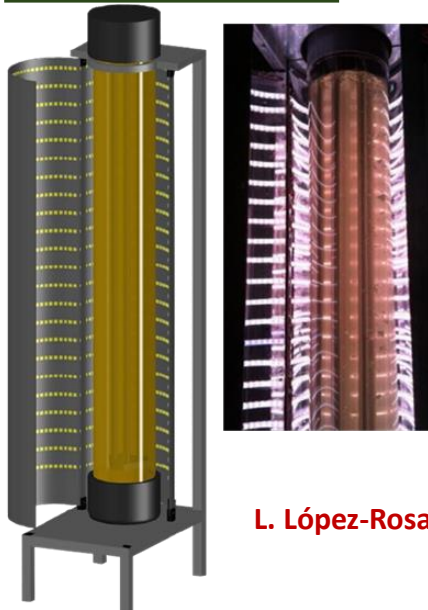
**López-Rosales, L.,** García-Camacho, F., Sánchez-Mirón, A., Contreras-Gómez, A., & Molina-Grima, E. (2017). Modeling shear-sensitive dinoflagellate microalgae growth in bubble column photobioreactors. *Bioresource technology*, 245, 250-257.

# Pilot-scale photobioreactors illuminated with LEDs

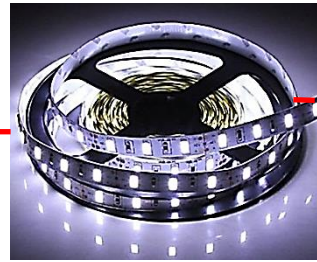
## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess**
4. Conclusions

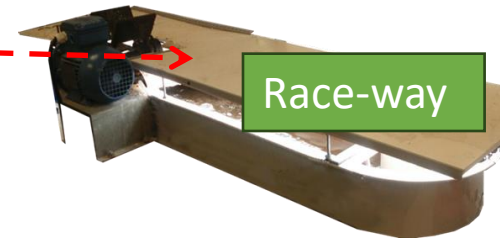
## Bubble column



L. López-Rosales et al., 2016

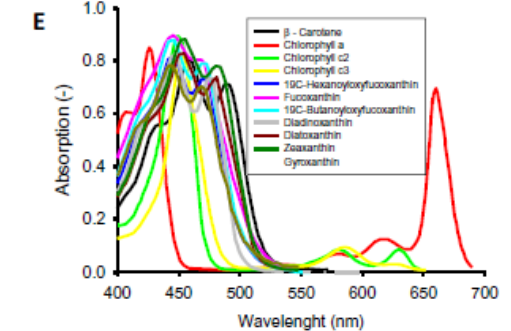
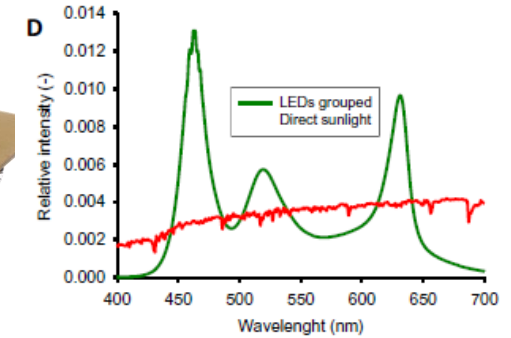


A. Molina-Miras et al., 2018b



## Race-way

Each LED was a multi-chip LED with the ability to provide multiple colors.



Absorption spectra of all the key pigments matches with the emission spectrum of the LEDs used. Therefore, LEDs had the potential to support algal photosynthesis

## Criteria of scale-up of bubble column and flat panel PBRs

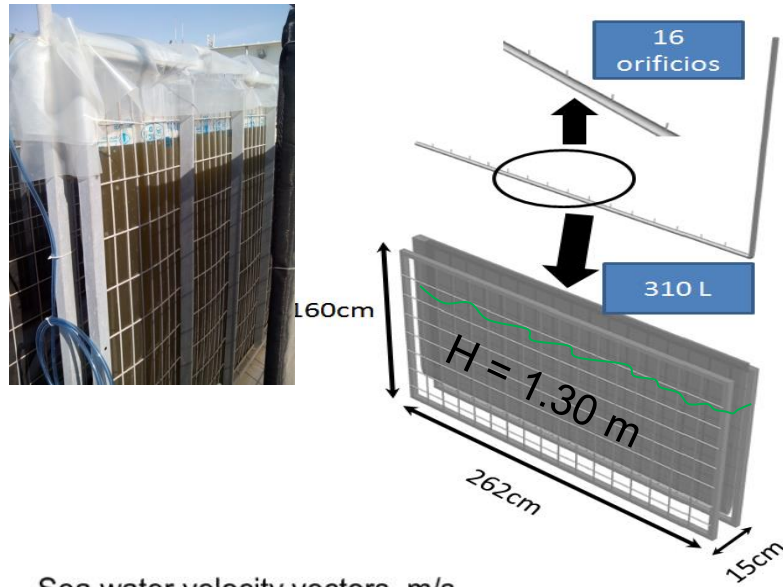


$H > 1.25$  m and  $d_c/d_o \approx 20$  assured freedom from damaging levels of hydrodynamic stress so long as the superficial aeration velocity ( $v_s$ ) remained below a species-dependent threshold value.

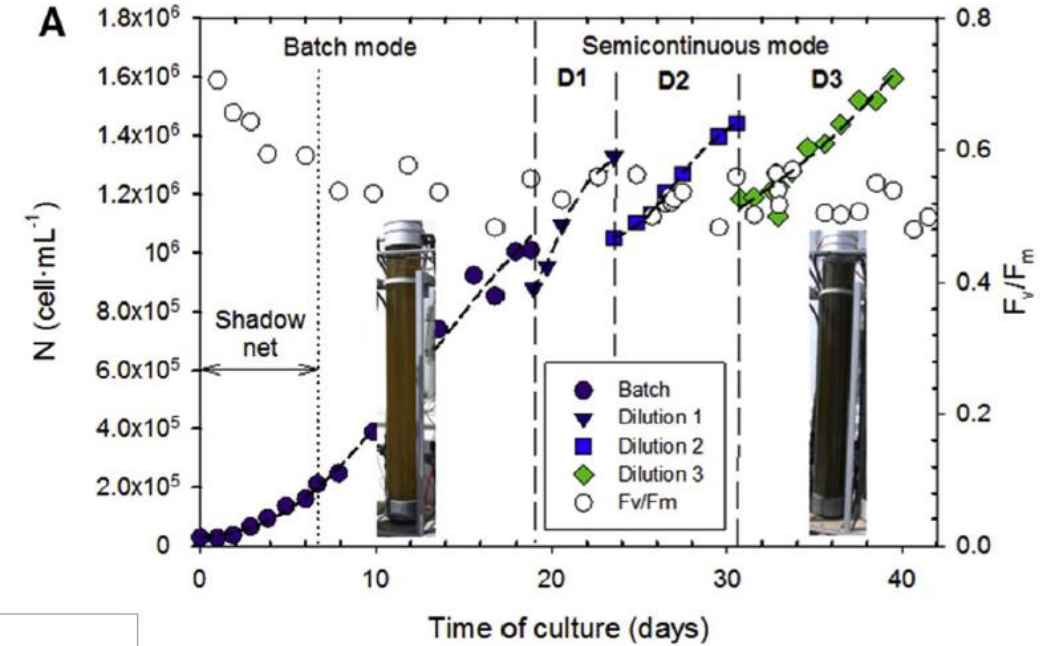
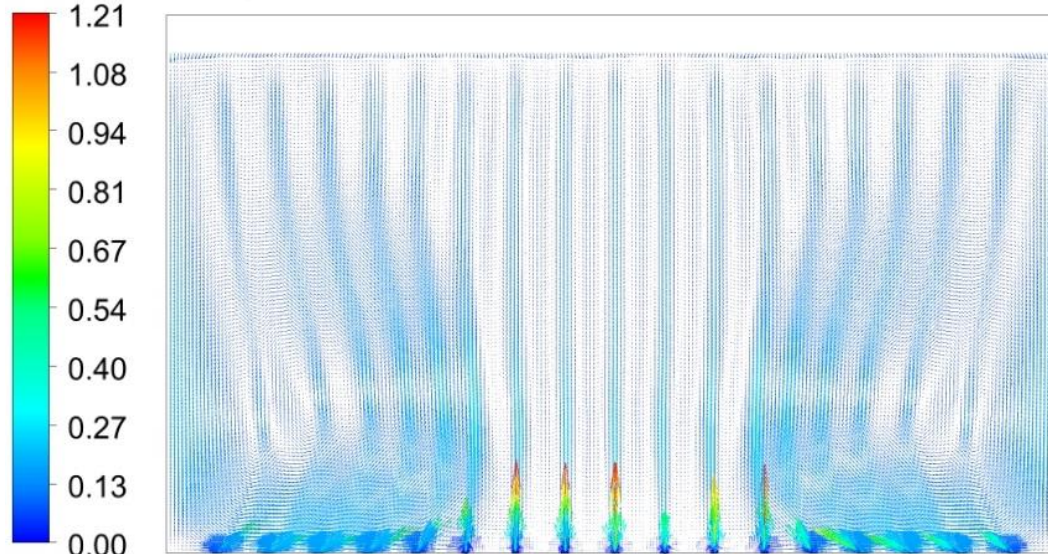
# Pilot-scale outdoor flat panel photobioreactor

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess**
4. Conclusions



Sea water velocity vectors, m/s



L. López-Rosales et al., 2018

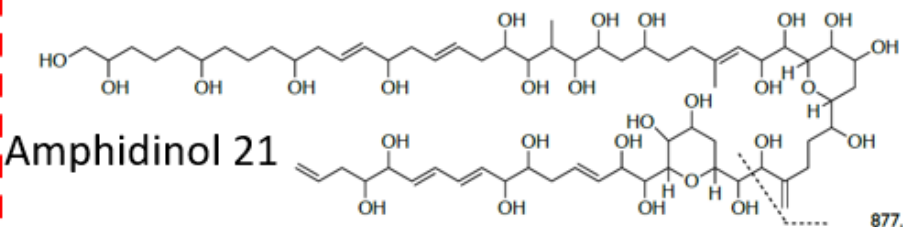
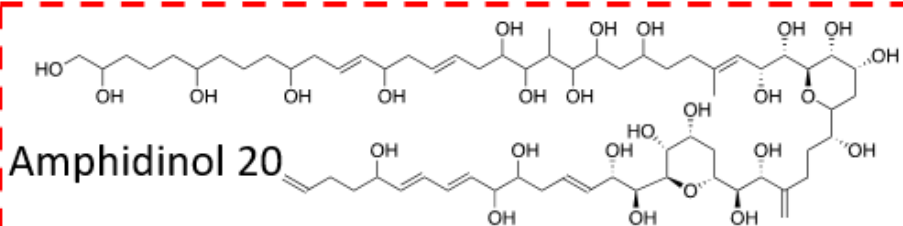
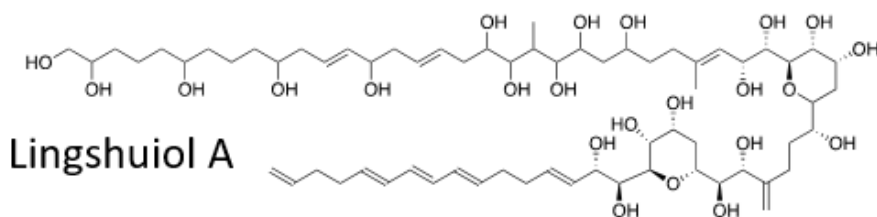
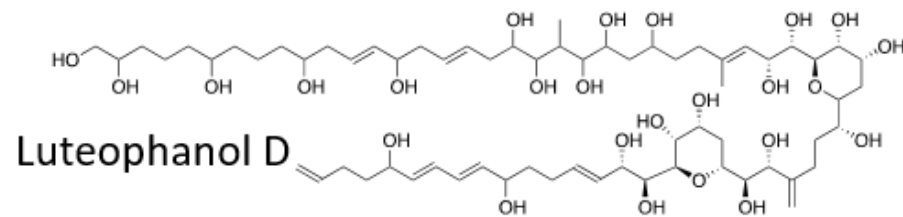
400L

# Some of the secondary metabolites isolated from our cultures so far

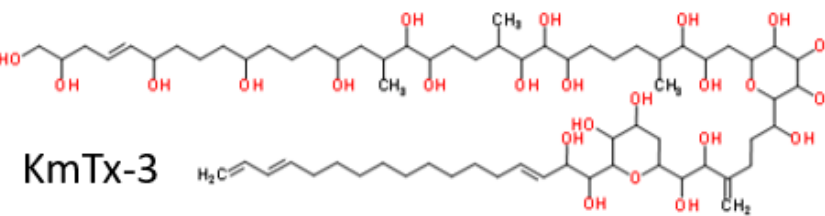
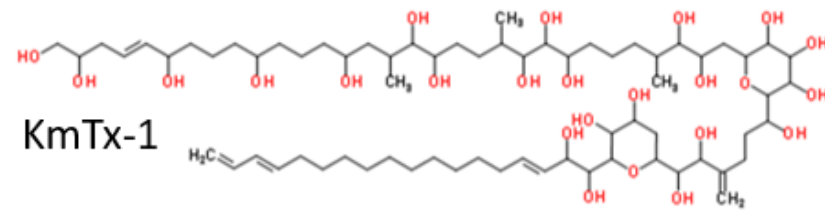
## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

### *Amphidinium carterae*



### *Karlodinium veneficum*

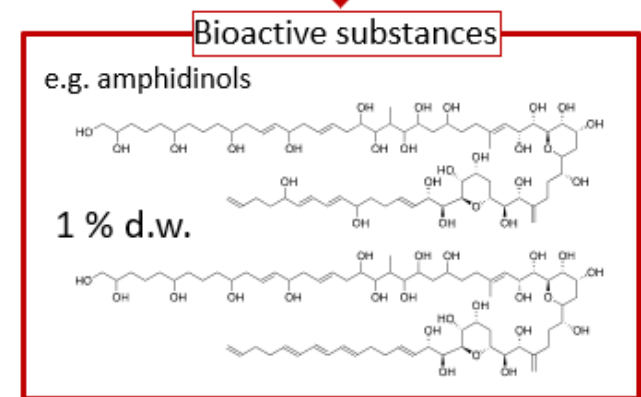
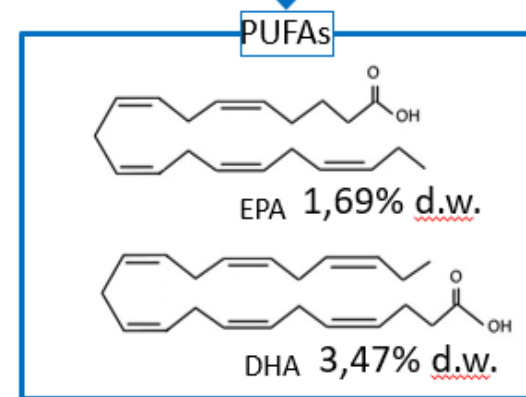
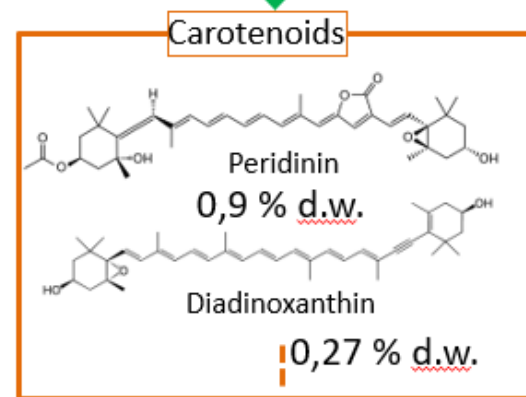
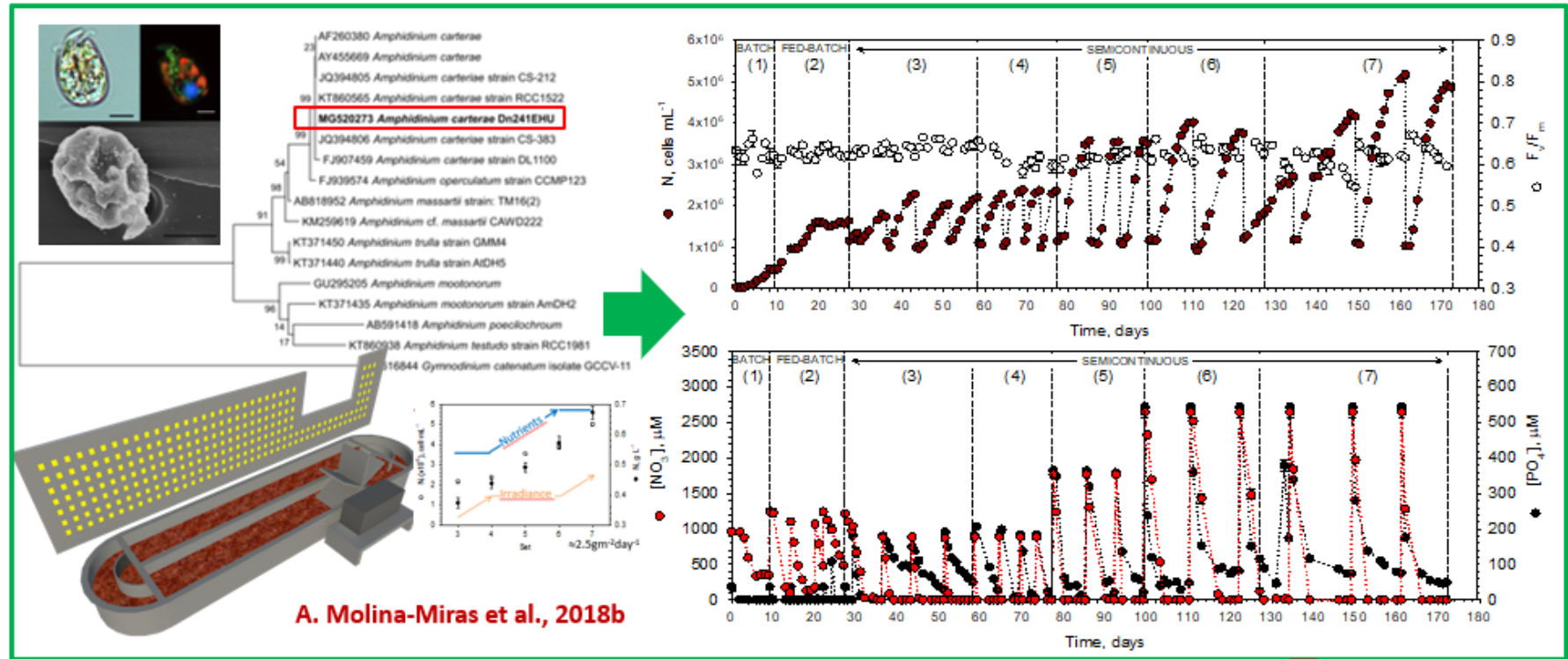


Compound	m/z	Elemental composition
KmTx-1303	1303.73	C <sub>62</sub> H <sub>117</sub> ClNaO <sub>22</sub> S
KmTx-1379	1379.75	C <sub>64</sub> H <sub>121</sub> ClNaO <sub>25</sub> S
KmTx-1423	1423.68	C <sub>62</sub> H <sub>118</sub> ClNa <sub>2</sub> O <sub>26</sub> S <sub>2</sub>
KmTx-1321	1321.74	C <sub>62</sub> H <sub>119</sub> ClNaO <sub>23</sub> S
KmTx-1355	1355.75	C <sub>62</sub> H <sub>121</sub> ClNaO <sub>25</sub> S

# Example of *A. carterae* Dn241EHU , RW-PBR

## OUTLINE

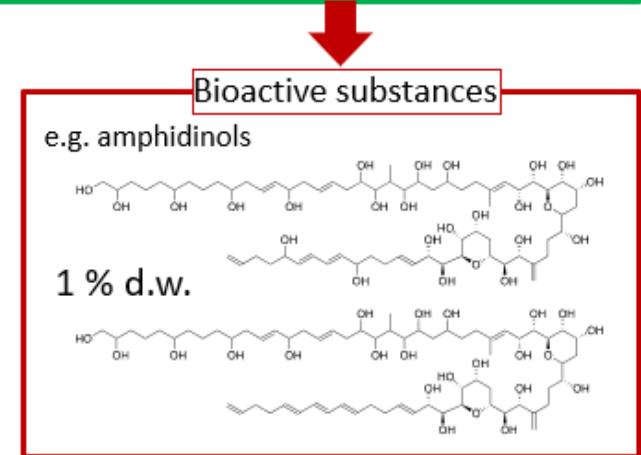
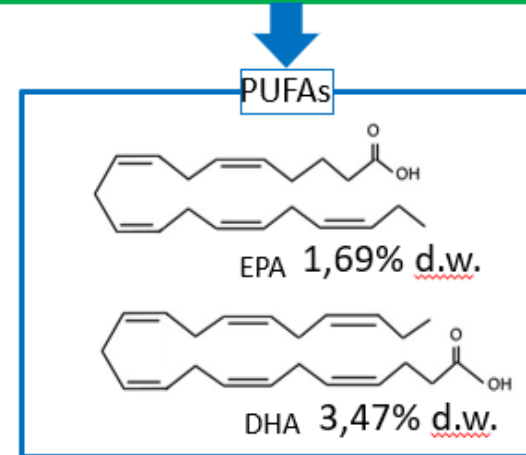
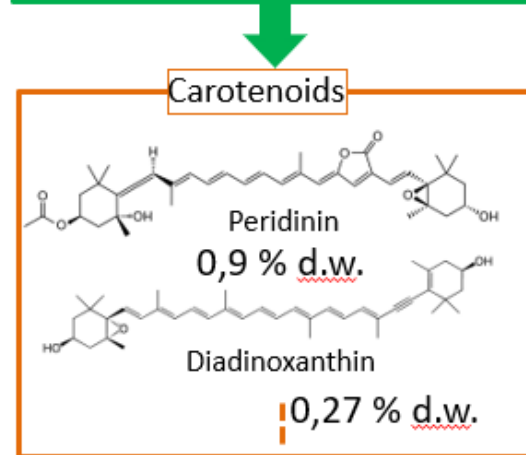
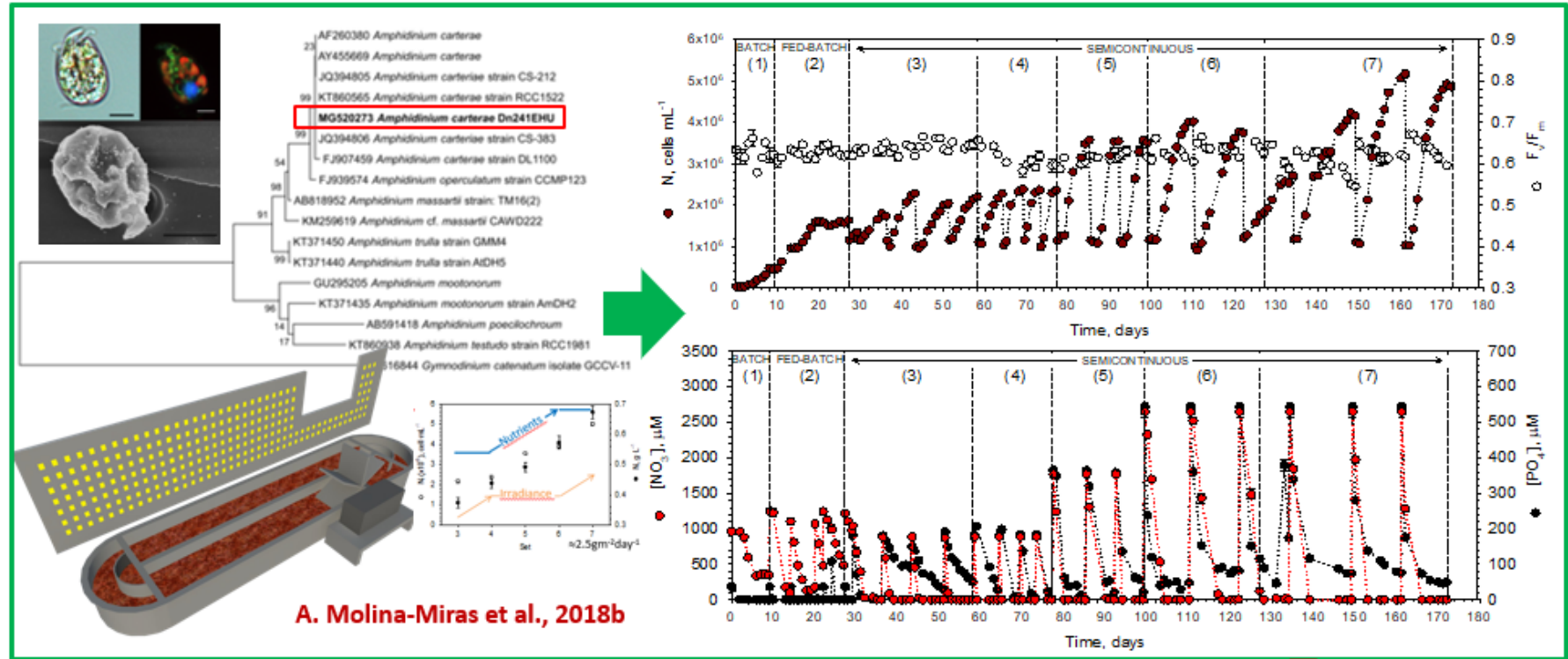
1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



# Example of *A. carterae* Dn241EHU , RW-PBR

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions



# 4. Conclusions

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
- 4. Conclusions**



# Conclusions

## OUTLINE

1. Overview
2. Objectives
3. Strategies
  - 3.1 Improvement of culture media
  - 3.2 Modelling the growth of dinoflagellates with ANNS
  - 3.3 Optimization of culture conditions
  - 3.4 Recycling recourses
  - 3.5 Scale-up of the bioprocess
4. Conclusions

- The use of genetic algorithms (**GAs**) has been shown as an **efficient strategy for developing complex culture media and optimization of culture conditions** in bubble columns
- The use of artificial neural networks (**ANNs**) in **modeling the growth of dinoflagellates** allowed to determine **the relative importance of nutrient medium on growth**
- **Optimization** of small-scale **culture conditions** is necessary to study the growth and production of biocompounds. Due to the **high sensitivity** to hydrodynamic stress of dinoflagellates, it is necessary to pay attention to the **operating conditions of the PBR.**
- **Water treatment** bioprocesses can be adapted to cultivate dinoflagellates and obtain high added value products.
- The **scale-up** of bioprocesses based on dinoflagellates **is possible. Robust and stable cultures can be achieved..**